

RESEARCH ARTICLE

Open Access



MiRNA155HG polymorphisms influenced the risk of liver cancer among the Han Chinese population

Xu Chao^{1,2*}, Xuesong Feng^{1†}, Xiaoping Wang¹, Hailong Shi¹, Hong Li², Yuewen Wang¹, Lanlan Wang¹, Haiyu Shen¹, Qing Zha¹ and Yanni Chen¹

Abstract

Background: Liver cancer is one of the most common cancers in the world. The primary aim of this research was to discover the correlation between single nucleotide polymorphisms (SNPs) of the *MIR155HG* and liver cancer risk.

Methods: The selected SNPs in *MIR155HG* were genotyped utilizing the Agena MassARRAY platform. We evaluated the correlation between *MIR155HG* polymorphisms and Liver cancer by genetic model analysis, stratification analysis and haplotype analysis. Relative risk of Liver cancer was shown based on odds ratios (ORs) and 95% confidence intervals (95% CIs).

Results: Our results uncovered that rs12482371 and rs1893650 in the *MIR155HG* were associated with protection against Liver cancer. And the rs928883 was related to increase risk of Liver cancer. Furthermore, apart from rs77218221, other selected SNPs formed two LD blocks, and haplotype "GATAG" in block 2 elevated individual liver cancer risk.

Conclusions: *MIR155HG* gene polymorphism may be correlated to Liver cancer susceptibility in Han Chinese population, particularly in males and aged ≤ 55 years.

Keywords: Liver cancer, *MIR155HG*, Single nucleotide polymorphisms (SNPs), Case-control study

Background

Globally, Liver cancer is a common malignancy and the second leading cause of cancer death worldwide [1], accounting for 466,100 new patients and 422,100 deaths per year in China [2]. The major type of Liver cancer (70–80%) is hepatocellular carcinoma (HCC), followed by intrahepatic cholangio carcinoma [3]. Among all cancers in China, liver cancer had poorest survival with a 5-year relative survival rate of only 10.1% [4]. Due to the difficulty of early diagnosis, most of the patients are

identified at an advanced stage with numerous lesions and extrahepatic metastasis. Moreover, as a highly aggressive malignancy with rapid progression, Liver cancer is difficult to control because of low selectivity and limited chemotherapeutic drugs [5]. So there is urgent need to better understand the potential mechanism of cancer development, especially in Chinese people. Liver cancer is a complex process, associated with many factors and co-factors. Hepatitis B virus (HBV) is commonly accepted to contribute to be the leading cause of Liver cancer [6, 7]. Occupational and environmental factors, such as certain metabolic Liver diseases and cirrhosis, have also been reported to contribute to the incidence of Liver cancer [8]. In addition, similarly with the breast cancer and glioma [9, 10], the occurrence of Liver cancer

* Correspondence: chaoxu2004@126.com

[†]Xu Chao and Xuesong Feng are Joint first author.

¹The College of Basic Medicine, The Shaanxi University of Chinese Medicine, Xianyang 712046, Shaanxi, China

²The Second Affiliated Hospital of Shaanxi University of Chinese Medicine, Xianyang 712000, Shaanxi, China



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

is also influenced by genetic factors, and the associations have been proved in many studies [11–13]. For example, Kumar V et al. found that genetic variant rs2596542 located in the 5' flanking region of MICA was strongly linked to HCV-induced hepatocellular carcinoma [14]. Zhang H et al. identified that internal polymorphism locus rs17401966 in KIF1B gene was highly correlated with HBV-associated HCC [15].

Long noncoding RNAs (lncRNAs) have no coding protein potential with a length of more than 200 nucleotides. They have been mistaken for genetic dark matter for a long time. However, in recent years, studies have confirmed that such RNAs participate in a variety of regulation in biological activities, such as transcriptional inhibition and post-transcriptional regulation [16]. *MIR155HG* is a typical lncRNA, known as B-cell Integration Cluster (*BIC*) [17]. This gene located on chromosome 21 of human. Tam subsequently proved that it is encoded in a highly conserved region of the third exon of the *BIC* [18]. In addition, miR-155 expression is closely related to tumor. Xuechao Wu et al. revealed that *MIR155HG* can promote the development of glioma, and showed cancer-promoting activity by expressing miR-155-5p/-3p [19]. Siqi Chen et al. discovered that host miR-155 activates tumor growth via a myeloid-derived mechanism of inhibiting cell-dependence [20]. Given the importance of *MIR155HG* in glioma, we hypothesized that germline genetic variants within *MIR155HG* would influence the course of liver cancer. As the most common genetic variation, single nucleotide polymorphisms (SNPs) can influence the development of diseases by influencing gene expression and its function. Therefore, we conducted a case-control study to discover the effects of genetic polymorphisms in *MIR155HG* on liver cancer susceptibility in Han Chinese population.

Methods

Study participants

A total of 432 liver cancer cases were recruited from the Shaanxi University of Chinese Medicine from August 2017 to December 2018. The inclusion criteria for eligible case were newly diagnosed primary liver cancer patients and histologically confirmed by at least two pathologists. The exclusion criteria for patients were family history of non-liver cancer, or any other malignancy and digestive diseases, such as hepatitis and cirrhosis of the liver. None of the patients received any treatment before blood collection. During the same period, 430 healthy controls were randomly enrolled from the same hospital's physical examination center. The exclusion criteria of controls were family history of liver cancer more than three generations and chronic digestive disease. All participants were Han Chinese and there was no blood relationship between each participant.

DNA extraction

Blood samples from each participant were drawn and stored with ethylenediamine tetraacetic acid (EDTA) at -80°C . Genomic DNA was extracted from peripheral blood using the GoldMag extraction method (GoldMag Co Ltd., Xi'an, China). The quality of DNA samples was detected by the NanoDrop 2000 (Thermo Scientific, Waltham, Massachusetts, USA).

SNPs selection and genotyping

The genetic information of the SNPs was obtained from dbSNP database and 1000 Genomes database (<http://www.internationalgenome.org/>). Only the variant with minor allele frequency (MAFs) $> 5\%$ in Asian population are eligible in this study in order to achieve adequate statistical power. Therefore, we finally picked up eight candidate polymorphisms (rs4143370, rs77218221, rs12482371, rs77699734, rs11911469, rs1893650, rs34904192, and rs928883) to perform analysis in Chinese Han population. According to Agena online software, the primers for the eight related sites were designed (Supplementary Table 1). SNPs were genotyped by Agena MassARRAY RS1000 (Agena, San Diego, CA, U.S.A.) (Supplementary Table 2) [21].

Statistical analysis

We applied SPSS version 19.0 (SPSS, Chicago, IL, USA) and Microsoft Excel software for statistical analysis. Hardy-Weinberg equilibrium in controls was determined via χ^2 test [22]. The distribution differences were evaluated using the chi-square test for sex and independent sample student's t test for age between cases and controls. The odds ratio (OR) values and 95% confidence intervals (CIs) were calculated to assess the relationships between *MIR155HG* polymorphisms and liver cancer risk [23]. The influence of SNPs in the *MIR155HG* on individual liver cancer risk was assessed by four genetic model analyses (genotype, dominant, recessive and log-additive) through PLINK software (Version 1.07). Further, we conducted stratification analysis by both age and sex. Then, statistical power of each SNP was examined through the online software (<http://sampsize.sourceforge.net/iface/s3.html#ccp>) in our research. Finally, the Haploview software package (version 4.2) was used to assess the linkage disequilibrium (LD) and performed haplotype analysis.

Results

Characteristics of the participants

We selected 432 liver cancer cases and 430 healthy controls in this research. The case group included 344 males and 88 females. The control group included 342 males and 88 females. The average age among cases and controls was 55.09 ± 11.59 years and 55.22 ± 10.73 years, respectively. The demographic characteristics of the case

and control groups were listed in Table 1. The statistical analysis results manifested that age and gender between two groups had no statistically significant difference ($p > .05$).

The associations between *MIR155HG* and liver cancer

Basic information of candidate SNPs in the *MIR155HG* were listed in Table 2, including chromosomal position, allele, minimum allele frequency (MAF), HWE test results and the p value of allele model analysis. All the selected SNPs met HWE ($p > 0.05$). We evaluated the risk of gene polymorphism in the allele model by chi-squared test, and found that there were no associations between the SNPs and the risk of Liver cancer with all $p > 0.05$.

As shown in Table 3, four genetic models (codominant, dominant, recessive, and additive genetic models) were applied to analyze the correlation between SNPs and liver cancer. For each variation, the minor allele was a risk factor when compared to the wild-type (major) allele in the genetic model. Our results found that rs12482371 in *MIR155HG* was associated with a 0.56-fold decrease in risk for Liver cancer under genotype model (adjusted, OR = 0.56, $p = 0.022$ for the “C/C” genotype) and a 0.55-fold decreased in risk for Liver cancer under recessive model (adjusted, OR = 0.55, $p = 0.015$ for the “C/C” genotype). The rs1893650 in *MIR155HG* was associated with a 0.35-fold decrease in risk for liver cancer under genotype model (adjusted, OR = 0.35, $p = 0.030$ for the “C/C” genotype) and a 0.34-fold decreased in risk for Liver cancer under recessive model (adjusted, OR = 0.34, $p = 0.026$ for the “C/C” genotype). The rs928883 in *MIR155HG* was associated with a 1.71-fold increase in risk for Liver cancer under genotype model (adjusted, OR = 1.71, $p = 0.001$ for the “A/G” genotype) and a 1.60-fold increase in risk for Liver cancer under dominant model (adjusted, OR = 1.60, $p = 0.003$ for the “A/G-A/A” genotype). We have found no other SNPs on *MIR155HG* associated with the

susceptibility to Liver cancer risk, either uncorrected or adjusted for age and sex.

Statistical power of rs12482371, rs1893650, rs928883 was estimated through the online software (<http://sample-size.sourceforge.net/iface/s3.html#ccp>), which were related to liver cancer risk in our research. The powers for rs12482371, rs1893650, rs928883 were 68.97, 63.95, and 97.12%, achieved a certain statistical power.

Stratification analysis of age and sex

Next, stratification analysis by age or gender was carried out to evaluate the influence of these candidate SNPs on liver cancer risk (Table 4). The stratification analysis of sex revealed the associations between three SNPs and Liver cancer risk. Two loci (rs12482371 and rs1893650) decreasing risk of Liver cancer in male under genotype model (rs12482371, CC vs T/T, OR = 0.51, $p = 0.016$; rs1893650, CC vs TT, OR = 0.32, $p = 0.030$) and recessive model (rs12482371, C/C vs T/T-C/T, OR = 0.52, $p = 0.018$; rs1893650, CC vs T/T-C/T, OR = 0.32, $p = 0.030$). Rs928883 was observed to increase Liver cancer risk under genotype model (AG vs GG, OR = 1.95, $p = 0.000$) and dominant model (AG-AA vs GG, OR = 1.74, $p = 0.002$). No statistical correlations were found in female in any genetic model.

Furthermore, we conducted the stratification analysis of age. At age > 55 years group and age ≤ 55 years group, we found that *MIR155HG* rs928883 added Liver cancer risk under genotype model (AG vs GG: age > 55 years group: OR = 1.84, $p = 0.013$; age ≤ 55 years group: OR = 1.62, $p = 0.036$) and dominant model (AG-AA vs GG: age > 55 years group: OR = 1.63, $p = 0.036$; age ≤ 55 years group: OR = 1.59, $p = 0.034$). For *MIR155HG* rs1893650, we only found that *MIR155HG* rs1893650 reduced Liver cancer risk in age ≤ 55 years under genotype model (CC vs TT, OR = 0.10, $p = 0.030$) and recessive model (CC vs TT-CT, OR = 0.10, $p = 0.029$).

Haplotype analyses

LD construction and haplotype analyses of these SNPs were performed in cases and controls. All SNPs existed in two LD blocks (Block 1: rs4143370 and rs12482371; Block 2: rs77699734, rs11911469, rs1893650, rs34904192 and rs928883) in *MIR155HG* (Fig. 1). Haplotype “GATAG” in Block 2 increased the risk of liver cancer (OR = 1.44, $p = 0.047$) (Table 5). However, we didn't find a significant correlation between the other haplotypes and Liver cancer risk.

Discussion

In this study, we assessed the associations of selected SNPs in *MIR155HG* with liver cancer susceptibility in the Han Chinese population. We demonstrated that *MIR155HG* genetic polymorphisms are correlated with

Table 1 Characteristics of patients with liver cancer and controls

Characteristics	Cases <i>n</i> = 432	Controls <i>n</i> = 430	<i>p</i> -value
Age, years	55.09 ± 11.59	55.22 ± 10.73	0.972 ^a
> 55	209	185	
≤ 55	223	245	
Gender			0.861 ^b
Male	344	342	
Female	88	88	

^a*p* values were calculated by Student's *t* tests

^b*p* values were calculated from two-sided chi-square tests

Table 2 Basic information about the candidate SNPs in this study

SNP ID	Chr.Position	Alleles (minor/major)	MAF		<i>p</i> - HWE	OR (95%CI)	<i>p</i> - value
			Cases	Controls			
rs4143370	21:25564661	C/G	0.169	0.154	0.715	1.12 (0.86 - 1.44)	0.407
rs77218221	21:25565063	C/T	0.047	0.046	0.613	1.02 (0.66 - 1.60)	0.918
rs12482371	21:25566041	C/T	0.284	0.317	0.095	0.85 (0.69 - 1.05)	0.125
rs77699734	21:25566995	C/G	0.074	0.095	0.784	0.76 (0.54 - 1.07)	0.120
rs11911469	21:25567971	A/C	0.122	0.107	0.804	1.16 (0.86 - 1.56)	0.334
rs1893650	21:25568503	C/T	0.182	0.196	0.879	0.91 (0.72 - 1.16)	0.447
rs34904192	21:25569623	A/G	0.238	0.248	0.438	0.95 (0.76 - 1.18)	0.634
rs928883	21:25571713	A/G	0.494	0.454	0.624	1.18 (0.97 - 1.42)	0.099

CI confidence interval, HWE Hardy–Weinberg equilibrium, MAF minor allele frequency; *p*-HWE < 0.01 indicates imbalance, OR odds ratio, SNP single-nucleotide polymorphism

**p* < 0.05 indicates statistical significance

**p* < 0.00625 (0.05/8) indicates statistical significance for Bonferroni correction

Table 3 Relationships between *MIR155HG* polymorphism and liver cancer risk

SNP ID	Model	Genotype	Case (number)	Control (number)	Before adjusted		After adjusted		Power	
					OR (95%CI)	<i>p</i> ^a	OR (95%CI)	<i>p</i> ^b		
rs12482371	Genotype	TT	217	208	1.00		1.00		68.97%	
		CT	185	171	1.04 (0.78 - 1.38)	0.801	1.04 (0.78 - 1.37)	0.801		
		CC	30	51	0.56 (0.35 - 0.92)	0.022*	0.56 (0.35 - 0.92)	0.022*		
	Dominant	TT	217	208	1.00		1.00			
		CT-CC	215	222	0.93 (0.71 - 1.21)	0.585	0.93 (0.71 - 1.21)	0.586		
	Recessive	TT-CT	402	379	1.00		1.00			
CC		30	51	0.55 (0.35 - 0.89)	0.014*	0.55 (0.35 - 0.89)	0.015*			
rs1893650	Log-additive	–	–	–	0.85 (0.70 - 1.05)	0.129	0.85 (0.70 - 1.05)	0.129	63.95%	
		Genotype	TT	281	279	1.00		1.00		
			CT	145	135	1.07 (0.80 - 1.42)	0.661	1.07 (0.80 - 1.42)		0.663
	CC		6	17	0.35 (0.14 - 0.90)	0.030*	0.35 (0.14 - 0.90)	0.030*		
	Dominant	TT	281	279	1.00		1.00			
		CT-CC	151	152	0.99 (0.75 - 1.30)	0.923	0.99 (0.75 - 1.3)	0.922		
Recessive	TT-CT	426	414	1.00		1.00				
	CC	6	17	0.34 (0.13 - 0.88)	0.026*	0.34 (0.13 - 0.88)	0.026*			
rs928883	Log-additive	–	–	–	0.91 (0.71 - 1.16)	0.433	0.90 (0.71 - 1.16)	0.432	97.12%	
		Genotype	GG	90	129	1.00		1.00		
			AG	244	204	1.71 (1.24 - 2.38)	0.001*	1.71 (1.24 - 2.38)		0.001*
	AA		85	90	1.35 (0.91 - 2.02)	0.138	1.35 (0.9 - 2.02)	0.141		
	Dominant	GG	90	129	1.00		1.00			
		AG-AA	329	294	1.60 (1.17 - 2.19)	0.003*	1.60 (1.17 - 2.19)	0.003*		
Recessive	GG-AG	334	333	1.00		1.00				
	AA	85	90	0.94 (0.67 - 1.31)	0.723	0.94 (0.67 - 1.31)	0.716			
Log-additive	–	–	–	–	1.19 (0.97 - 1.45)	0.088	1.19 (0.97 - 1.45)	0.090		

OR odds ratio, SNP single-nucleotide polymorphism, 95% CI 95% confidence interval. ^a*p* values were calculated from unconditional logistic regression analysis. ^b*p* values were adjusted by age and gender

**p* < 0.05 indicates statistical significance

**p* < 0.0002 (0.05/8/4) indicates statistical significance for Bonferroni correction

Table 4 The relationship of three gene polymorphisms with liver cancer according to the stratification by gender and age

SNP	Model	Genotype	Male		Female		> 55		≤55	
			OR (95%CI)	p	OR (95%CI)	p	OR (95%CI)	p	OR (95%CI)	p
rs12482371	Allele	T	1.00		1.00		1.00		1.00	
		C	0.79 (0.63 - 1.00)	0.046	1.11 (0.71 - 1.76)	0.642	0.88 (0.65 - 1.19)	0.410	0.81 (0.61 - 1.08)	0.155
	Genotype	TT	1.00		1.00		1.00		1.00	
		CT	0.93 (0.68 - 1.28)	0.651	1.55 (0.83 - 2.92)	0.170	1.08 (0.71 - 1.65)	0.718	0.97 (0.66 - 1.43)	0.895
	Dominant	CC	0.51 (0.29 - 0.88)	0.016*	0.82 (0.29 - 2.36)	0.716	0.60 (0.30 - 1.21)	0.156	0.52 (0.26 - 1.04)	0.063
		CT-CC	0.83 (0.62 - 1.13)	0.236	1.38 (0.76 - 2.49)	0.293	0.97 (0.65 - 1.44)	0.871	0.87 (0.60 - 1.26)	0.460
	Recessive	TT-CT	1.00		1.00		1.00		1.00	
		CC	0.52 (0.31 - 0.90)	0.018*	0.67 (0.24 - 1.86)	0.448	0.58 (0.30 - 1.14)	0.112	0.52 (0.27 - 1.03)	0.059
	Log-additive	-	0.79 (0.63 - 1.00)	0.049	1.11 (0.71 - 1.74)	0.648	0.88 (0.65 - 1.19)	0.395	0.82 (0.62 - 1.08)	0.161
	rs1893650	Allele	T	1.00		1.00		1.00		1.00
C			0.85 (0.65 - 1.11)	0.224	1.22 (0.70 - 2.12)	0.481	1.01 (0.71 - 1.44)	0.956	0.82 (0.59 - 1.15)	0.252
Genotype		TT	1.00		1.00		1.00		1.00	
		CT	0.99 (0.71 - 1.36)	0.929	1.43 (0.75 - 2.72)	0.279	1.13 (0.74 - 1.73)	0.577	1.00 (0.67 - 1.48)	0.984
Dominant		CC	0.32 (0.11 - 0.90)	0.030*	0.55 (0.05 - 6.25)	0.627	0.66 (0.20 - 2.14)	0.491	0.10 (0.01 - 0.80)	0.030*
		CT-CC	0.91 (0.66 - 1.24)	0.537	1.36 (0.72 - 2.56)	0.339	1.08 (0.71 - 1.63)	0.719	0.89 (0.61 - 1.32)	0.570
Recessive		TT-CT	1.00		1.00		1.00		1.00	
		CC	0.32 (0.12 - 0.90)	0.030*	0.49 (0.04 - 5.51)	0.560	0.63 (0.20 - 2.04)	0.445	0.10 (0.01 - 0.79)	0.029*
Log-additive		-	0.84 (0.64-1.11)	0.212	1.24 (0.69 - 2.23)	0.464	1.01 (0.71 - 1.46)	0.941	0.80 (0.56 - 1.13)	0.206
rs928883		Allele	G	1.00		1.00		1.00		1.00
	A		1.17 (0.94 - 1.45)	0.155	1.20 (0.78-1.83)	0.406	1.12 (0.85 - 1.49)	0.420	1.22 (0.94 - 1.59)	0.130
	Genotype	GG	1.00		1.00		1.00		1.00	
		AG	1.95 (1.34 - 2.82)	0.000*	1.11 (0.55 - 2.23)	0.766	1.84 (1.14 - 2.98)	0.013*	1.62 (1.03 - 2.55)	0.036*
	Dominant	AA	1.34 (0.86 - 2.09)	0.203	1.54 (0.62 - 3.87)	0.356	1.20 (0.66 - 2.17)	0.547	1.53 (0.88 - 2.65)	0.130
		GG-AA	1.74 (1.23 - 2.48)	0.002*	1.20 (0.62 - 2.34)	0.588	1.63 (1.03 - 2.58)	0.036*	1.59 (1.04 - 2.45)	0.034*
	Recessive	GG-AG	1.00		1.00		1.00		1.00	
		AA	0.85 (0.59 - 1.23)	0.399	1.44 (0.65 - 3.19)	0.370	0.80 (0.49 - 1.31)	0.373	1.10 (0.70 - 1.74)	0.671
	Log-additive	-	1.18 (0.95 - 1.47)	0.144	1.22 (0.78 - 1.92)	0.381	1.13 (0.84 - 1.52)	0.408	1.25 (0.95 - 1.65)	0.106

OR odds ratio, 95%CI 95% confidence interval

Bold indicates statistical significance ($p < 0.05$)* $p < 0.00625$ (0.05/8) indicates statistical significance for Bonferroni correction

liver cancer risk among Han Chinese individuals. Our findings uncovered that rs12482371 and rs1893650 in *MIR155HG* were related to reduce Liver cancer risk, and rs928883 increased Liver cancer risk. Furthermore, apart from the rs77218221, other selected SNPs could form two LD blocks. Haplotype “GATAG” in Block 2 was found to elevate the liver cancer risk. These results indicated the importance of *MIR155HG* gene polymorphism in liver cancer development among the Han Chinese population.

Non-coding RNAs are non-protein encoding transcripts closely related to gene transcription, epigenetic regulation

and protein translation, which include miRNAs, piwiRNAs and long non-coding RNAs (lncRNAs). Previous studies have confirmed the functional interaction between lncRNA and miRNA, as well as the fact that some lncRNAs themselves may encode miRNAs [24]. The dis-regulation of miR-155 can affect cancer development [25]. The increased expression of miR-155 accompanied by the occurrence of several cancers such as glioma, breast cancer, lung cancer and gastric cancer has been established recently [26, 27]. Therefore, targeting miR-155 is considered a promising method for the treatment of hematopoietic and solid tumors. *MIR155HG* was the primary micro (mi)RNA of miR-

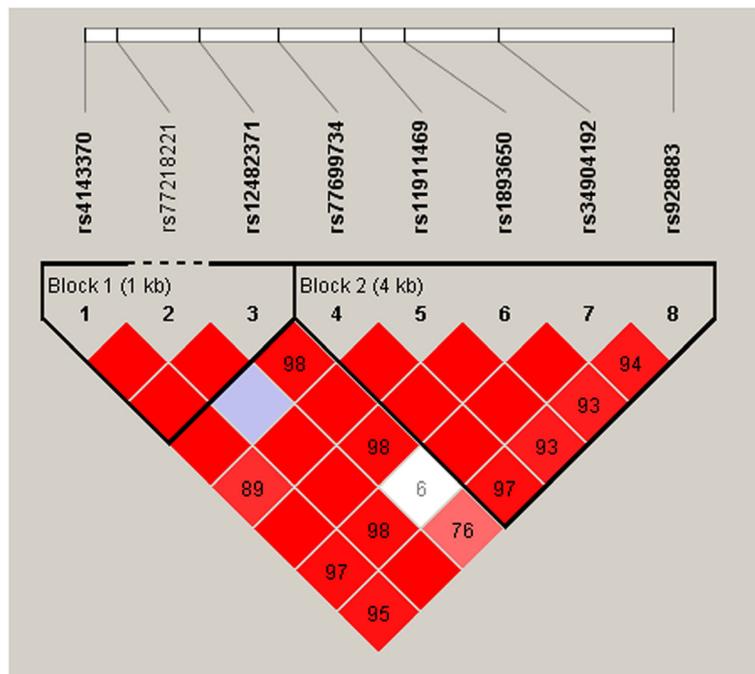


Fig. 1 Linkage disequilibrium (LD) plots containing SNPs from *MIR155HG*

155 [18]. However, underlying mechanisms of host miR-155 in modulating tumor growth are still poorly understood. Xuechao Wu et al. demonstrated that *MIR155HG* was an independent adverse prognostic factor in glioma. And its repression inhibited glioma growth in vitro and in vivo [19]. Veerakumar Balasubramaniyan et al. raised that targeting *MIR155HG* can be a novel approach in glioma [28]. And miR-155 is a transcriptional product of its host gene (*MIR155HG*), the genetic variation of *MIR155HG* gene and *miR-155* gene may affected its expression [29].

To date, however, there were no studies focusing on the influence of *MIR155HG* genetic variations on liver cancer risk. Our findings suggest that our hypothesis that *MIR155HG* polymorphisms were related to liver cancer risk in Chinese is correct. Our results showed that rs12482371 and rs1893650 in the *MIR155HG* were associated with protection from Liver cancer. And the rs928883 was connected with an added risk of Liver cancer. One research found that the allele “C” of rs12482371 and rs1893650, and the allele “A” of

Table 5 Haplotype analysis results in this study

Block	Haplotype	Frequency		OR (95%CI)	p
		Case	Control		
Block 1	rs4143370 rs12482371				
	GT	0.284	0.317	0.85 (0.70 - 1.05)	0.129
	CC	0.831	0.845	0.9 (0.70 - 1.16)	0.424
	GC	0.548	0.528	1.08 (0.90 - 1.31)	0.414
Block 2	rs77699734 rs11911469 rs1893650 rs34904192 rs928883				
	CATGA	0.527	0.554	0.9 (0.74 - 1.09)	0.267
	GATAG	0.932	0.905	1.44 (1.01 - 2.06)	0.047*
	CATAG	0.850	0.849	1.01 (0.78 - 1.32)	0.932
	CACGG	0.172	0.190	0.87 (0.68 - 1.13)	0.311
	CCTGG	0.116	0.106	1.12 (0.82 - 1.52)	0.477

OR odds ratio, *pa* adjusted by gender and age, *SNP* single nucleotide polymorphism, *95% CI* 95% confidence interval
 Bold indicates statistical significance (*p* < 0.05)

rs928883 in *MIR155HG* were correlated to colorectal cancer risk [30]. Moreover, the sites selected in this study are existed in the intron region of *MIR155HG*. This is consistent with our recent understanding of introns. Introns are no longer what we traditionally think of as meaningless. Studies have shown that it can influence transcription level by affecting transcription rate, nuclear output and transcription stability, and also can improve the efficiency of mRNA translation [31]. Although the specific mechanism of rs12482371, rs1893650, and rs928883 on the *MIR155HG* affecting liver cancer is unclear, it is worthwhile to reveal the correlation between the SNPs and liver cancer susceptibility. Meanwhile, our results will be helpful for the prevention and early detection of Liver cancer and give theoretical basis.

There are several limitations to consider in the current study. Our research is fundamental, and it does not performed experiments to investigate the expression of miR-155 and its predicted targets. So, the exact molecular mechanism by which SNP alters the expression of *MIR155HG* needs to be clarified in future work.

Conclusions

To sum up, our research first demonstrated that *MIR155HG* gene polymorphisms were correlated with the susceptibility to Liver cancer among Han Chinese population, in particular males and age ≤ 55 years old.

Supplementary information

Supplementary information accompanies this paper at <https://doi.org/10.1186/s12881-020-01064-4>.

Additional file 1: Table 1. PCR primer for this study

Additional file 2: Table 2. the Genetic polymorphisms of eight locus in 432 liver cancer cases and 430 healthy controls

Abbreviations

SNPs: Single nucleotide polymorphisms; miRNA-155 HG: miRNA-155 host gene; OR: Odds ratio; CI: Confidence interval; MAF: Minor allele frequency; EDTA: Ethylenediamine tetraacetic acid; HWE: Hardy–Weinberg equilibrium; LD: Linkage disequilibrium; HCC: Hepatocellular carcinoma; HBV: Hepatitis B virus; LncRNAs: Long noncoding RNAs

Acknowledgements

We thank all of the participants for their involvement in this study. We are very grateful to the clinicians and other hospital staff for providing blood samples and data collection for this study.

Authors' contributions

XC conceived and designed the experiments. XPW, HLS and YNC performed genotyping experiment. HL and YWW extracted the DNA. LLW, HYS, and QZ completed the statistical analysis. XC and XSF wrote the manuscript. All authors approved the final manuscript.

Funding

This research was supported by the National Natural Science Foundation of China (No. 81774132) and Subject Innovation Team of the Shaanxi University of Chinese Medicine (No. 2019-YS05), which helped us in financial.

Availability of data and materials

All data generated or analysed during this study are included in this published article and its supplementary information files.

Ethics approval and consent to participate

This study was conducted in compliance with the Declaration of Helsinki, and approved by the Research Ethics Committee of the Shaanxi University of Chinese Medicine. Written informed consent was obtained from all participants before taking part in this research. The design and performance of this study involving human subjects were implemented in accordance with the approved guidelines.

Consent for publication

Not Applicable.

Competing interests

None.

Received: 28 June 2019 Accepted: 5 June 2020

Published online: 19 June 2020

References

- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin*. 2015;65(2):87–108.
- Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ, He J. Cancer statistics in China, 2015. *CA Cancer J Clin*. 2016;66(2):115–32.
- de Jong MC, Nathan H, Sotiropoulos GC, Paul A, Alexandrescu S, Marques H, Pulitano C, Barroso E, Clary BM, Aldrighetti L, et al. Intrahepatic cholangiocarcinoma: an international multi-institutional analysis of prognostic factors and lymph node assessment. *J Clin Oncol*. 2011;29(23):3140–5.
- Zeng H, Zheng R, Guo Y, Zhang S, Zou X, Wang N, Zhang L, Tang J, Chen J, Wei K, et al. Cancer survival in China, 2003–2005: a population-based study. *Int J Cancer*. 2015;136(8):1921–30.
- Fu J, Wang H. Precision diagnosis and treatment of liver cancer in China. *Cancer Lett*. 2018;412:283–8.
- Levero M, Zucman-Rossi J. Mechanisms of HBV-induced hepatocellular carcinoma. *J Hepatol*. 2016;64(1 Suppl):S84–s101.
- Wen J, Song C, Jiang D, Jin T, Dai J, Zhu L, An J, Liu Y, Ma S, Qin N, et al. Hepatitis B virus genotype, mutations, human leukocyte antigen polymorphisms and their interactions in hepatocellular carcinoma: a multi-Centre case-control study. *Sci Rep*. 2015;5:16489.
- McGlynn KA, Petrick JL, London WT. Global epidemiology of hepatocellular carcinoma: an emphasis on demographic and regional variability. *Clin Liver Dis*. 2015;19(2):223–38.
- Li G, Zhang Z, Tu Y, Jin T, Liang H, Cui G, He S, Gao G. Correlation of microRNA-372 upregulation with poor prognosis in human glioma. *Diagn Pathol*. 2013;8:1.
- Xia P, Li B, Geng T, Deng Z, Dang C, Chang D, Kang L, Jin T, Chen C. FGFR2 gene polymorphisms are associated with breast cancer risk in the Han Chinese population. *Am J Cancer Res*. 2015;5(5):1854–61.
- Li Y, Sun X, Zhan SH, Gao YQ, Xin YN, Xuan SY. [research progress of single-nucleotide polymorphism in the TM6SF2 rs58542926]. *Zhonghua Gan Zang Bing Za Zhi*. 2019;27(3):223–6.
- Yadegar A, Mohabati Mobarez A, Zali MR. Genetic diversity and amino acid sequence polymorphism in helicobacter pylori CagL hypervariable motif and its association with virulence markers and gastroduodenal diseases. *Cancer Med*. 2019;8(4):1619–32.
- Dondeti MF, El-Maadawy EA, Talaat RM. Hepatitis-related hepatocellular carcinoma: insights into cytokine gene polymorphisms. *World J Gastroenterol*. 2016;22(30):6800–16.
- Kumar V, Kato N, Urabe Y, Takahashi A, Muroyama R, Hosono N, Otsuka M, Tateishi R, Omata M, Nakagawa H, et al. Genome-wide association study identifies a susceptibility locus for HCV-induced hepatocellular carcinoma. *Nat Genet*. 2011;43(5):455–8.
- Zhang H, Zhai Y, Hu Z, Wu C, Qian J, Jia W, Ma F, Huang W, Yu L, Yue W, et al. Genome-wide association study identifies 1p36.22 as a new susceptibility locus for hepatocellular carcinoma in chronic hepatitis B virus carriers. *Nat Genet*. 2010;42(9):755–8.
- Yoon JH, Abdelmohsen K, Gorospe M. Posttranscriptional gene regulation by long noncoding RNA. *J Mol Biol*. 2013;425(19):3723–30.
- Tam W, Ben-Yehuda D, Hayward WS. bic, a novel gene activated by proviral insertions in avian leukosis virus-induced lymphomas, is likely to function through its noncoding RNA. *Mol Cell Biol*. 1997;17(3):1490–502.

18. Tam W. Identification and characterization of human BIC, a gene on chromosome 21 that encodes a noncoding RNA. *Gene*. 2001;274(1-2):157–67.
19. Wu X, Wang Y, Yu T, Nie E, Hu Q, Wu W, Zhi T, Jiang K, Wang X, Lu X, et al. Blocking MIR155HG/miR-155 axis inhibits mesenchymal transition in glioma. *Neuro Oncol*. 2017;19(9):1195–205.
20. Yu F, Jia X, Du F, Wang J, Wang Y, Ai W, Fan D: miR-155-deficient bone marrow promotes tumor metastasis. *Mol Cancer Res*. 2013;11(8):923–36.
21. Gabriel S, Ziaugra L, Tabbaa D. SNP genotyping using the Sequenom MassARRAY iPLEX platform. *Curr Protoc Hum Genet*. 2009;2:2–12.
22. Adamec C. Example of the use of the nonparametric test. Test X2 for comparison of 2 independent examples. *Cesk Zdrav*. 1964;12:613–9.
23. Bland JM, Altman DG. Statistics notes. The odds ratio. *BMJ*. 2000;320(7247):1468.
24. Briggs JA, Wolvetang EJ, Mattick JS, Rinn JL, Barry G. Mechanisms of long non-coding RNAs in mammalian nervous system development, plasticity, disease, and evolution. *Neuron*. 2015;88(5):861–77.
25. Tili E, Croce CM, Michaille JJ: miR-155: on the crosstalk between inflammation and cancer. *Int Rev Immunol*. 2009;28(5):264–84.
26. O'Connell RM, Rao DS, Chaudhuri AA, Baltimore D. Physiological and pathological roles for microRNAs in the immune system. *Nat Rev Immunol*. 2010;10(2):111–22.
27. Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, Visone R, Iorio M, Roldo C, Ferracin M, et al. A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci U S A*. 2006; 103(7):2257–61.
28. Balasubramanian V, Bhat KP. Targeting MIR155HG in glioma: a novel approach. *Neuro Oncol*. 2017;19(9):1152–3.
29. Tao H, Cui L, Li Y, Zhou X, Ma G, Yao L, Fu J, Li W, Cai Y, Zhou H, et al. Association of tag SNPs and rare CNVs of the MIR155HG/miR-155 gene with epilepsy in the Chinese Han population. *Biomed Res Int*. 2015;2015:837213.
30. Wu H, He G, Han H, Xiong W, Song T, Chen H, Chen X, Wu X, Huang G, Zhang Y, et al. Analysis of MIR155HG variants and colorectal cancer susceptibility in Han Chinese population. *Mol Genet Genom Med*. 2019;7(8):e778.
31. Shaul O. How introns enhance gene expression. *Int J Biochem Cell Biol*. 2017;91(Pt B):145–55.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

