

CASE REPORT

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Identification of a novel *GLA* mutation (Y88C) in a Korean family with Fabry nephropathy: a case report

Yosep Chong¹, Minyoung Kim², Eun Sil Koh², Seok Joon Shin², Ho-Shik Kim³ and Sungjin Chung^{2*} 

Abstract

Background: Fabry disease is a rare X-linked lysosomal storage disorder caused by α -galactosidase A deficiency. With the advancement of molecular diagnostic tools, more disease-causing mutations in α -galactosidase A (*GLA*) have been identified in Fabry disease. We found a novel mutation in a Korean family with predominant renal manifestations of the disease.

Case presentation: A 24-year-old man who wanted to donate a kidney to his 28-year-old brother with end-stage renal disease of unknown cause was evaluated. The 24-year-old man underwent percutaneous renal biopsy because of an accidentally found proteinuria. Electron microscopy of his renal biopsy showed numerous electron-dense multi-lamellar inclusions in the epithelial cytoplasm, typical for Fabry disease. Clinical and laboratory evaluation including the assessment of *GLA* enzyme activity and direct DNA sequencing in four members of the family were performed. Renal biopsy findings in the two affected male patients were described. Re-evaluation of a renal biopsy specimen of his 28-year-old brother obtained when he was diagnosed with renal failure revealed a very focal area of suspicious multilamellated structures in the Bowman's space. DNA sequencing on the young man, his brother, and his mother revealed a novel *GLA* gene mutation, c.263A > G (p.Tyr88Cys). The three all showed decreased α -galactosidase A activity.

Conclusion: A novel *GLA* mutation, c.263A > G (p.Tyr88Cys), was found in a Korean family with predominant renal manifestations of Fabry disease.

Keywords: α -galactosidase A, Dialysis, Fabry disease, Kidney biopsy, Proteinuria

Background

Fabry disease (FD, OMIM#301500) is a rare, progressive, X-linked, and multisystemic disorder characterized by α -galactosidase deficiency resulting from mutations in the α -galactosidase (*GLA*, OMIM*300644) gene at Xq22.1 [1, 2]. The enzyme α -galactosidase A (*GLA*) is a ubiquitous lysosomal acid hydrolase. Its deficient activity can lead to progressive accumulation of globotriaosylceramide and other glycosphingolipids within the lysosomes of various cell types throughout the body, causing severe and potentially life-threatening target-organ complications such as cardiovascular diseases, stroke, renal failure, and eventually death [2].

The spectrum of clinical presentations in male patients with FD varies widely, from severe classic phenotype to relatively mild late-onset variant form [2, 3]. According to previous reports, the clinical manifestations in the classic form of FD starting from childhood or adolescence include severe symptoms such as acroparesthesia, hypohidrosis, corneal opacities, stroke, cardiac abnormalities, and renal disorders with high mortality [3–5]. On the contrary, the clinical manifestations of the variant form comprising almost 70 % of FD patients [6] are usually mild and limited to the heart or kidney [3, 4]. Due to mild symptoms, these patients are often found after disease progression. They only show findings of focal segmental glomerular sclerosis. For such cases, the key pathognomonic features such as electron-dense multi-lamellar inclusions can be obscured by extensive sclerosis on renal biopsy [4, 6]. As a result, it can be difficult to diagnose FD in time, particularly its variant

* Correspondence: sungjin.chung@outlook.com

²Department of Internal Medicine, College of Medicine, The Catholic University of Korea, Seoul, Republic of Korea

Full list of author information is available at the end of the article



form which is often undiagnosed or misdiagnosed as other heart or kidney diseases [4].

Recently, there have been amazing advances in molecular diagnostics for FD. More than 700 mutations in *GLA* gene have been identified, including missense mutations, small deletions/insertions, splice mutations, and large gene rearrangements [7]. These are listed in the Human Gene Mutation Database (<http://www.hgmd.cf.ac.uk/>), Fabry Database (<http://fabry-database.org/>), and ClinVar (<http://www.ncbi.nlm.nih.gov/clinvar/>). New mutations are still being discovered.

In the present study, we identified a novel mutation, Tyr88Cys, in *GLA* in a Korean family with late-onset FD variant. We also discussed their clinicopathologic features and the molecular diagnosis of FD.

Case presentation

Subjects and kidney biopsy procedure

We studied four members of a Korean family. Clinical and physical examinations were performed for each member. All family members gave written consent before we obtained blood and kidney samples from them.

Percutaneous renal biopsy was done by nephrologists under ultrasonographic guidance using an automated biopsy gun as previously described [8]. Histopathologic diagnosis on each sample was made comprehensively based on all clinical data and pathologic findings.

Analysis of genomic DNA and GLA enzyme activity

Genomic DNA samples were isolated from peripheral white blood cells using QIAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany). For sequencing, samples were pooled in an equimolar ratio. Genomic DNA library (10 pM) containing 1 % PhiX control library were prepared and sequenced on a MiSeq platform (Illumina, San Diego, CA) using MiSeq v2 chemistry. Next generation sequencing (NGS) data analyses were performed using bioinformatics pipeline of Labgenomics Co. Ltd. (Gyeonggi-do, Republic of Korea). Briefly, the paired-end sequencing reads of 250 bases in length were binned for each sample according to the index and mapped to the human genome assembly (hg19). Burrows-Wheeler Aligner (BWA) was used to align these sequences to the

human genome allowing up to 2 base mismatches (RefSeq NM_000169). Pathogenic variants and variants of unknown significance (VUS) detected by NGS were confirmed by Sanger sequencing analyses with AB 3730XL DNA Analyser (Life Technologies, Carlsbad, CA, USA) in accordance with the manufacturer's instructions. Sequencing results were searched against data of Korean exomes obtained from Labgenomics, data from Korea National Institution of Health (KNHIIH), dbSNP, and 1000 Genomes Project databases. To determine whether the newly discovered mutation in *GLA* might contribute to FD, prediction tests including Polyphen-2 and MutationTaster were used as described previously [9–11].

For all subjects, we assessed lysosomal *GLA* enzyme in the leukocytes by fluorometric assay using substrate 4-MU- α -D-galactopyranoside [12, 13]. Enzyme level was expressed as nmole of cleaved substrate per milligram of protein per hour at 37 °C. The enzyme activity was normalized to the activity of the control (100 %).

Results

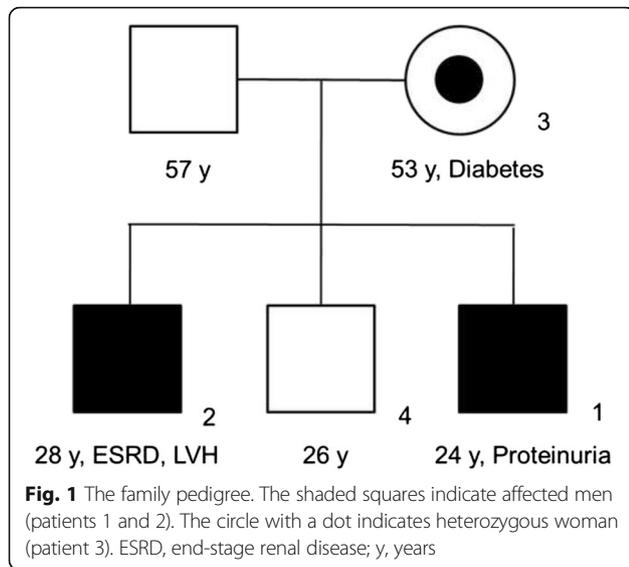
The clinicopathologic features of these patients including enzymatic and genetic analysis results are summarized in Table 1. The pedigree of the family is shown in Fig. 1. A 24-year-old man (patient 1) was evaluated voluntarily to donate a kidney to his 28-year-old brother (patient 2) who had end-stage renal disease (ESRD) and had been undergoing peritoneal dialysis for the previous 2 years. Patient 1 had an unremarkable medical history without medications. His assessment was overall satisfactory except a proteinuria of 871 mg/day (reference range, < 150 mg/day) that was discovered incidentally. A kidney biopsy was performed in view of the unaccountable proteinuria.

In 2014, which was two years ago before patient 1's visit, patient 2 (his brother) had presented at the age of 26 years with nephrotic-range proteinuria (24 h urine protein, 8486.4 mg/day) and renal insufficiency with serum creatinine of 8.06 mg/dL (reference, 0.50–1.20 mg/dL). At that time, a renal biopsy was performed and the finding was glomerulosclerosis with moderate to severe

Table 1 Clinicopathologic features of the four patients described in this case report

Subject No.	Age/Sex	Clinical findings	α -galactosidase A	c.263A > G mutation	Kidney biopsy findings
Patient 1	24/M	Proteinuria	Decreased (2.2 nmol/hr/mg protein)	Present	Consistent with FD
Patient 2	28/M	End-stage renal disease, hypertension, left ventricular hypertrophy	Decreased (4.0 nmol/hr/mg protein)	Present	Consistent with FD
Patient 3	54/F	Mild proteinuria, diabetes	Slightly decreased (32.6 nmol/hr/mg protein)	Present	Unavailable
Individual 4	26/M	Unremarkable	Normal (43.4 nmol/hr/mg protein)	Not present	Unavailable

FD Fabry disease, F female, M male, No, number



tubulointerstitial fibrosis indicating advanced renal failure. As a result, plans to initiate dialysis were immediately made. Considering patient 1's proteinuria, there was ample room for doubt regarding familial kidney disease. Accordingly, patient 2's previous renal biopsy slides and medical records were re-reviewed.

The mother (patient 3) of patients 1 and 2 was a 53-year-old female who had been diagnosed with diabetes 10 years earlier. She had been completely asymptomatic. However, based on her sons' results, she also underwent screening. Results revealed normal renal function with slightly elevated proteinuria (154.8 mg/day). Renal biopsy in patient 3 was not performed. Individual 4, the second son of patient 3, showed normal results for all laboratory findings, including renal function.

Apart from renal involvement, the affected patients (1, 2, and 3) had no other clinical manifestations such as peripheral neuropathy, angiokeratoma of skin, corneal opacities, abdominal pain, diarrhea, or arrhythmias. Patient 2 only showed high blood pressure (165/100 mmHg) and left ventricular hypertrophy on echocardiographic examination.

Renal biopsy from patient 1 revealed 12 glomeruli with only a subtle change on microscopic examination. However, nothing was significant (Fig. 2a). The glomerular epithelial cells were swollen and showed somewhat bubbly appearance with vague vacuoles. The renal tubules, interstitium, and blood vessels were unremarkable. There was no immune deposition on immunofluorescence for immunoglobulins. Electron microscopy revealed typical electron-dense multi-lamellar inclusions and zebra bodies in the cytoplasm of the epithelial cells (Fig. 2b).

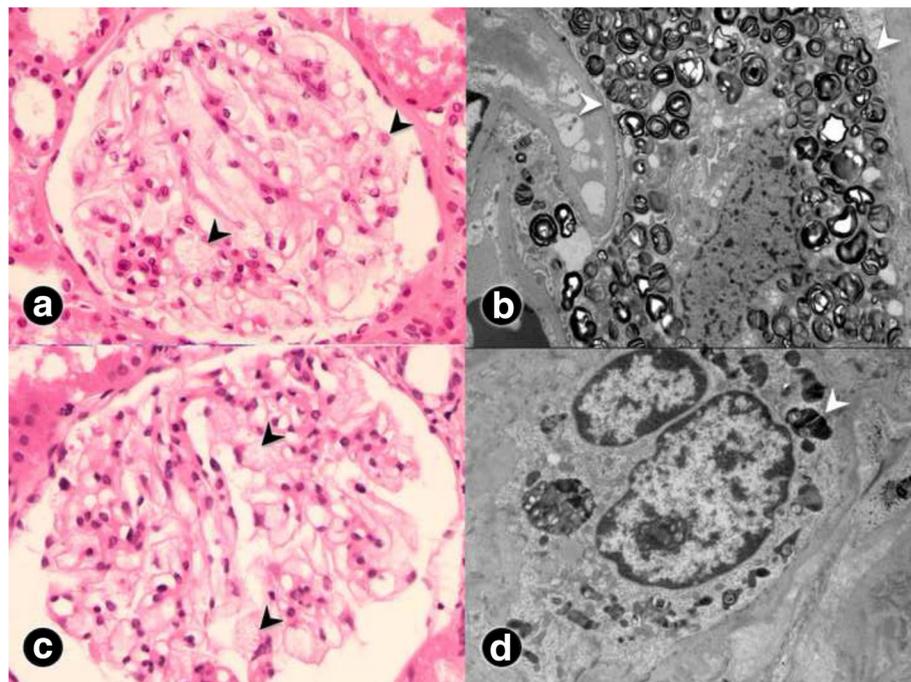


Fig. 2 Light and electron microscopic findings of renal biopsies from patients 1 and 2. **a** On microscopic examination of patient 1, there are some vague vacuolizations of podocytes (black arrowhead), although he showed normal renal function and proteinuria (H&E stain, $\times 400$). **b** Multilamellated myelin figures (white arrowhead), so-called zebra bodies—which are typical findings in Fabry disease—are found in patient 1's podocytes on electron microscopy. **c** Similar vacuolated epithelial cells (black arrowhead) are found in the relatively preserved glomerulus of patient 2, whereas most glomeruli are damaged by sclerosis (H&E stain, $\times 400$). **d** A focal area showing a tiny, dark-stained, round multilamellar inclusion (white arrowhead) on the electron microscopy result for patient 2

Patient 3 was thought to be a healthy female carrier. However, her GLA protein level and activity were 32.6 nmol/h/mg protein (average of 1,000 normal controls, 79.0 nmol/h/mg) and 89.6 % (average of 1,000 normal controls, 92.3 %), respectively, which were below normal ranges, indicating a heterozygous state. The protein level and activity of GLA enzyme in individual 4 were 43.4 nmol/h/mg protein (average of 1,000 normal controls, 61.6 nmol/h/mg) and 94.0 % (average of 1,000 normal controls, 93.2 %), respectively, which were within normal ranges.

Conclusion

In this report, we identified a new GLA mutation (c.263A > G p.Tyr99Cys) in a Korean family with predominant renal involvement of late-onset FD. We have deposited this new mutation data to the Fabry Database (<http://fabry-database.org/>, ID: 329). Although these family members shared the same GLA mutation, substantial differences in clinicopathologic features and residual enzyme activities were noticed, which can be potential pitfall for proper early diagnosis of FD, especially for its late-onset variant.

GLA mutation in coding sequence can result in amino acid changes, which is likely to alter protein conformation and subsequent functional defect of the GLA enzyme [14]. The 88 Tyr amino acid belongs to the group of amino acids buried in the hydrophobic core likely involved in protein folding [14, 15]. Although the current study did not cover the functional pathogenesis of Tyr88Cys, the potential deleterious effect of the Tyr88Cys mutation was predicted in this study using bioinformatics tools Polyphen-2 and MutationTaster. However, further study on the functional alteration resulted from Y88C mutation is still needed.

The genotype-phenotype correlation is not perfect in FD [5]. In this report, two brothers (patients 1 and 2) were only 4 years apart in age. However, they demonstrated somewhat different clinical courses. Patient 2 was already on peritoneal dialysis due to resultant ESRD while patient 1 only had mild proteinuria with normal renal function, although they had similar levels of GLA. This supports the notion that environmental and other genetic factors can affect the disease severity and phenotypic variability of FD [5]. This is consistent to the findings of a previous study showing marked intra-familial variability in residual GLA enzyme activity and the natural disease course of FD [16]. The GLA activity in patient 3, the mother, was slightly below the normal range without evident clinical symptoms. Female carriers of GLA mutations in FD have been considered as asymptomatic originally. However, it is now accepted that heterozygous females may also develop mild to severe clinical manifestations of FD [5, 17]. The

characteristic presentations including delayed onset and a higher probability of a single organ involvement have been thought to be due to random X chromosome inactivation [18, 19].

Recently found novel GLA mutations are mainly from patients with mild and late-onset FDs [13, 20]. Characteristically, late-onset FD has renal or cardiac involvement without characteristic skin lesions or pain crises. FD is often diagnosed when patients are in their 50 s or 60 s [11].

This report shows an important point regarding kidney biopsy. In patient 1 who had only proteinuria, biopsy played a valuable role in diagnosing the FD missed in his family member. Although his kidney specimen demonstrated almost normal appearance in light microscopy examination, the electron microscopy clearly demonstrated lamellated myelin structures and confirmed the diagnosis of typical FD. On the contrary, we missed the FD findings in patient 2's kidney biopsy 2 years ago due to extensive sclerotic glomerular damage. The lamellated structure covered on the glomerular surface may inhibit glomerular filtration and result in endothelial injury [15]. Subsequently, endothelial cells in the vicinity of any damaged podocytes are submitted to additional hydraulic forces, leading to glomerular collapse and sclerotic change [15]. This diffuse and progressive destruction of the kidney structure may make it difficult to recognize the typical FD findings. Thus, it is important to secure the minimum requirement of preserved glomeruli to obtain accurate pathologic diagnosis. For female patients 3 with minimal proteinuria, it might be necessary to perform renal biopsy in order to identify whether the proteinuria was due to FD or diabetes. The majority of female carriers do not develop renal disease with glycosphingolipids deposition. However, some clinically normal heterozygous females have typical kidney biopsy findings [21].

Early diagnosis of FD is crucial for commencing early enzyme replacement therapy and improving the natural disease course by preventing progressive organ failure [22]. Although diabetes, hypertension, and chronic glomerulonephritis are the most common causes of ESRD, the causes of primary renal disease in more than 20 % of patients with ESRD are still uncertain [23, 24]. Therefore, it is reasonable to bestow some consideration on screening for FD in patients with renal failure of unknown causes at an early age. In male patients, the diagnosis of FD can be made based on markedly deficient GLA enzyme activity in plasma or leukocytes and GLA mutations [9]. In order to identify affected females, it is important to combine biochemical assays with molecular analyses of the GLA gene [16, 17]. Considering that FD has progressive effects on various organs, vital organ damage would have already begun regardless of the

activity of GLA enzyme. This implies that biopsy of the involved organ could help us identify the extent of FD and start therapy accordingly.

Because the parents of patient 3 have already passed away with unspecified causes at the ages of 62 and 67 years, respectively, it is impossible to know at which generation that this mutation started in this family. However, patient 3 had seven siblings. Her two older brothers died of renal disease that required dialysis at the ages of 31 and 57 years, respectively. Molecular testing for *GLA* gene for patient 3's youngest brother was normal. Tests for other siblings are now being carried out. Past history of her two brothers suggests the possibility that the patient 3's mother was a carrier of the mutant *GLA* gene. However, it is also possible that this mutation is a *de novo* mutation or that either the mother or the father of patient 3 is a germ line mosaic for the mutation [25, 26]. In patients 1 and 2, the enzymatic activity of GLA needs to be tested serially so that treatment can be started before other organ involvement becomes irreversible. The female carrier patient 3 also needs to be followed up closely for the possibility of her developing FD-related symptoms. A request for enzyme replacement therapy has been submitted to The Health Insurance Review and Assessment Service.

In summary, this report revealed the presence of a c.263A > G p.Tyr88Cys mutation in exon 2 of *GLA* gene in a Korean family with Fabry nephropathy. The detection of this new mutation in *GLA* highlights the notion that there is no single predominant mutation in FD. This report also described the possibility that distinctive findings of Fabry nephropathy might not appear clearly in severely damaged kidneys.

Abbreviations

4-MU- α -D-galactopyranoside: 4-Methylumbelliferyl- α -D-galactopyranoside; Cys: Cysteine; dL: Deciliter; DNA: Deoxyribonucleic acid; ESRD: End-stage renal disease; FD: Fabry disease; GLA: α -galactosidase A; H&E: Hematoxylin and eosin; hr: Hour; kb: Kilobase; mg: Milligram; mmHg: Millimeters of mercury; nmol: Nanomole; Tyr: Tyrosine

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Availability of data and materials

All data supporting our findings are included in the manuscript.

Authors' contributions

SC, MK and ESK were involved in the care of patients. MK and ESK performed percutaneous renal biopsy of the patients. YC, SJS, H-SK and SC analyzed and interpreted the data. YC and SC prepared the manuscript. All authors read and approved the manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Written informed consents for publication of this Case report and any accompanying images were obtained from patients. A copy of the consent form is available for review by the Editor of this journal.

Ethics approval and consent to participate

This report was approved by the Institutional Review Board of The Catholic University of Korea Yeouido St. Mary's Hospital (SC16RISI0055). All analyses were done in accordance with the Helsinki Declaration of 1975.

Author details

¹Department of Hospital Pathology, College of Medicine, The Catholic University of Korea, Seoul, Republic of Korea. ²Department of Internal Medicine, College of Medicine, The Catholic University of Korea, Seoul, Republic of Korea. ³Department of Biochemistry, College of Medicine, The Catholic University of Korea, Seoul, Republic of Korea.

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References

- Echevarria L, Benistan K, Toussaint A, Dubourg O, Hagege AA, Eladiri D, et al. X-chromosome inactivation in female patients with Fabry disease. *Clin Genet*. 2016;89:44–54.
- Ortiz A, Abiose A, Bichet DG, Cabrera G, Charrow J, Germain DP, et al. Time to treatment benefit for adult patients with Fabry disease receiving agalsidase β : data from the Fabry Registry. *J Med Genet*. 2016 in press doi: 10.1136/jmedgenet-2015-103486.
- Nishino T, Obata Y, Furusu A, Hirose M, Shinzato K, Hattori K, et al. Identification of a novel mutation and prevalence study for fabry disease in Japanese dialysis patients. *Ren Fail*. 2012;34:566–70.
- Peng H, Xu X, Zhang L, Zhang X, Peng H, Zheng Y, et al. GLA variation p. E66Q identified as the genetic etiology of Fabry disease using exome sequencing. *Gene*. 2016;575:363–7.
- Zizzo C, Monte I, Pisani A, Fatuzzo P, Riccio E, Rodolico MS, et al. Molecular and clinical studies in five index cases with novel mutations in the GLA gene. *Gene*. 2016;578:100–4.
- Uribe A, Mateus HE, Prieto JC, Palacios MF, Ospina SY, Pasqualini G, et al. Identification of mutations in Colombian patients affected with Fabry disease. *Gene*. 2015;574:325–9.
- Tuttolomondo A, Duro G, Miceli S, Di Raimondo D, Pecoraro R, Serio A, et al. Novel alpha-galactosidase A mutation in a female with recurrent strokes. *Clin Biochem*. 2012;45:1525–30.
- Chung S, Koh ES, Kim SJ, Yoon HE, Park CW, Chang YS, et al. Safety and tissue yield for percutaneous native kidney biopsy according to practitioner and ultrasound technique. *BMC Nephrol*. 2014;15:96.
- Ng PC, Henikoff S. Predicting the effects of amino acid substitutions on protein function. *Annu Rev Genomics Hum Genet*. 2006;7:61–80.
- Schwarz JM, Cooper DN, Schuelke M, Seelow D. MutationTaster2: mutation prediction for the deepsequencing age. *Nat Methods*. 2014;11:361–2.
- Umeda T, Hashimoto S, Noriyasu K, Takamura A, Fujisaki M, Eto Y. Identification of a novel GLA mutation (F69 L) in a Japanese patient with late-onset Fabry disease. *Hum Genome Var*. 2015;2:15044.
- Han M, Jun SH, Song SH, Park KU, Kim JQ, Song J. Use of tandem mass spectrometry for newborn screening of 6 lysosomal storage disorders in a Korean population. *Korean J Lab Med*. 2011;31:250–6.
- Lukas J, Scalia S, Eichler S, Pockrandt AM, Dehn N, Cozma C, et al. Functional and Clinical Consequences of Novel α -Galactosidase A Mutations in Fabry Disease. *Hum Mutat*. 2016;37:43–51.
- Duro G, Musumeci MB, Colomba P, Zizzo C, Albeggiani G, Mastromarino V, et al. Novel α -galactosidase A mutation in patients with severe cardiac manifestations of Fabry disease. *Gene*. 2014;535:365–9.

15. Garman SC. Structure-function relationships in alpha-galactosidase A. *Acta Paediatr.* 2007;96:6–16.
16. Sessa A, Meroni M, Battini G, Righetti M, Nebuloni M, Tosoni A, et al. Studio Multicentrico Italiano sulla Malattia di Anderson-Fabry. Evolution of renal pathology in Fabry disease. *Acta Paediatr Suppl.* 2003;92:6–8.
17. Ferri L, Cavicchi C, Fiumara A, Parini R, Guerrini R, Morrone A. Pitfalls in the detection of gross gene rearrangements using MLPA in Fabry disease. *Clin Chim Acta.* 2016;452:82–6.
18. Yousef Z, Elliott PM, Cecchi F, Escoubet B, Linhart A, Monserrat L, Namdar M, Weidemann F. Left ventricular hypertrophy in Fabry disease: a practical approach to diagnosis. *Eur Heart J.* 2013;34:802–8.
19. Baptista A, Magalhães P, Leão S, Carvalho S, Mateus P, Moreira I. Screening for Fabry disease in left ventricular hypertrophy: documentation of a novel mutation. *Arq Bras Cardiol.* 2015;105:139–44.
20. Brouns R, Thijs V, Eyskens F, Van den Broeck M, Belachew S, Van Broeckhoven C, Redondo P, Hemelsoet D, Fumal A, Jeangette S, Verslegers W, Baker R, Hughes D, De Deyn PP, BeFaS Investigators. Belgian Fabry study: prevalence of Fabry disease in a cohort of 1000 young patients with cerebrovascular disease. *Stroke.* 2010;41:863–8.
21. Wüthrich RP, Weinreich T, Binswanger U, Gloor HJ, Candinas D, et al. Should living related kidney transplantation be considered for patients with renal failure due to Fabry's disease? *Nephrol Dial Transplant.* 1998;13:2934–6.
22. Okur I, Ezgu F, Biberoglu G, Tumer L, Erten Y, Isitman M, et al. Screening for Fabry disease in patients undergoing dialysis for chronic renal failure in Turkey: identification of new case with novel mutation. *Gene.* 2013;527:42–7.
23. Chung S, Chung JH, Kim SJ, Koh ES, Yoon HE, Park CW, et al. Blood lead and cadmium levels and renal function in Korean adults. *Clin Exp Nephrol.* 2014; 18:726–34.
24. Jin DC, Yun SR, Lee SW, Han SW, Kim W, Park J, et al. Lessons from 30 years' data of Korean endstage renal disease registry, 1985–2015. *Kidney Res Clin Pract.* 2015;34:132–9.
25. Pisani A, Daniele A, Di Domenico C, Nigro E, Salvatore F, Riccio E. Late diagnosis of Fabry disease caused by a de novo mutation in a patient with end stage renal disease. *BMC Res Notes.* 2015;8:711.
26. Iemolo F, Pizzo F, Albeggiani G, Zizzo C, Colomba P, Scalia S, Bartolotta C, Duro G. De novo mutation in a male patient with Fabry disease: a case report. *BMC Res Notes.* 2014;7:11.

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