

## **RESEARCH ARTICLE**

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# Associations of common polymorphisms in *GCKR* with type 2 diabetes and related traits in a Han Chinese population: a case-control study

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#### **Abstract**

**Background:** Several studies have shown that variants in the glucokinase regulatory protein gene (*GCKR*) were associated with type 2 diabetes and dyslipidemia. The purpose of this study was to examine whether tag single nucleotide polymorphisms (SNPs) in the *GCKR* region were associated with type 2 diabetes and related traits in a Han Chinese population and to identify the potential mechanisms underlying these associations.

**Methods:** We investigated the association of polymorphisms in the *GCKR* gene with type 2 diabetes by employing a case-control study design (1118 cases and 1161 controls). Four tag SNPs (rs8179206, rs2293572, rs3817588 and rs780094) with pairwise  $r^2 > 0.8$  and minor allele frequency > 0.05 across the *GCKR* gene and its flanking regions were studied and haplotypes were constructed. Genotyping was performed by matrix-assisted laser desorption/ionization time-of-flight mass spectroscopy using a MassARRAY platform.

**Results:** The G alleles of *GCKR* rs3817588 and rs780094 were associated with an increased risk of type 2 diabetes after adjustment for year of birth, sex and BMI (OR = 1.24, 95% CI 1.08-1.43, p = 0.002 and OR = 1.22, 95% CI 1.07-1.38, p = 0.002, respectively). In the non-diabetic controls, the GG carriers of rs3817588 and rs780094 were nominally associated with a lower plasma triglyceride level compared to the AA carriers after adjustment for year of birth, sex and BMI (p for trend = 0.00004 and 0.03, respectively). Furthermore, the association of rs3817588 with plasma triglyceride level was still significant after correcting for multiple testing.

**Conclusions:** The rs3817588 A/G polymorphism of the *GCKR* gene was associated with type 2 diabetes and plasma triglyceride level in the Han Chinese population.

#### **Background**

Glucokinase (GCK) is the key glucose phosphorylation enzyme responsible for the first rate-limiting step in the glycolysis pathway. GCK regulates glucose metabolism in the liver and glucose-stimulated insulin secretion from pancreatic beta cells [1]. GCK activity is closely regulated by the glucokinase regulatory protein (GCKR), a process depending on fructose 6-phosphate and fructose 1-phosphate [2,3]. Gckr-deficient mice display reduced GCK protein levels and activity in the liver and exhibit impaired postprandial glycemic control [4,5]. In a previous study, adenoviral-mediated hepatic overexpression of GCKR significantly improved insulin

sensitivity and glucose tolerance in mice and resulted in decreased leptin concentration and increased triglyceride levels [6].

In the Diabetes Genetics Initiative genome-wide association study, the GCKR rs780094 A allele was found to be strongly associated with hypertriglyceridemia in populations from Finland and Sweden [7]. Subsequently, a large study of Danish white participants confirmed that the rs780094 A allele was associated with increased fasting triglycerides, impaired fasting and OGTT-related insulin release, reduced homeostasis model assessment of insulin resistance (HOMA-IR), increased risk of dyslipidemia and a modestly decreased risk of type 2 diabetes [8]. The HapMap II CEU data http://www.hapmap.org showed that rs780094 was in strong linkage disequilibrium (LD) ( $r^2 = 0.932$ ) with a non-synonymous GCKR variant (Pro446Leu, rs1260326). The DESIR

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prospective cohort study demonstrated that the GCKR variant rs1260326 T allele was strongly associated with increased triglyceride levels, lower fasting glucose and insulin levels, a lower HOMA-IR index, and a higher risk for dyslipidemia, but a lower risk for hyperglycemia and type 2 diabetes in a general French population [9]. Another study, combining data from 12 independent cohorts comprising more than 45,000 individuals with various ethnic backgrounds, confirmed that GCKR rs780094 and rs1260326 were strongly associated with opposite effects on fasting plasma triglyceride and glucose concentrations [10]. Recently, the MAGIC study conducted a large-scale meta-analysis and provided convincing evidence that the GCKR rs780094 A allele was associated with lower fasting glucose and insulin levels, a lower HOMA-IR index, a higher triglyceride level, and a lower risk for type 2 diabetes [11].

Several studies of the association of GCKR variants with type 2 diabetes or glucose homeostasis parameters in Chinese populations have been reported [12-14]. In a study of a population-based sample of Han Chinese individuals, the GCKR rs780094 A allele was found to be significantly associated with a reduced risk of impaired fasting glucose (IFG) and type 2 diabetes, decreased fasting glucose, increased homeostasis model assessment of beta cell function (HOMA-B), and fasting triglyceride levels; GCKR rs1260326 displayed similar associations [12]. A study of healthy Chinese adults and adolescents showed that the GCKR rs780094 A allele was associated with increased triglyceride levels, and GCKR rs780094 alone did not contribute to fasting glucose but interacted with GCK rs1799884 to increase fasting glucose [13]. However, another study in a Han Chinese cohort did not find any association between GCKR rs780094 and type 2 diabetes [14]. Therefore, the association of GCKR variants with fasting plasma glucose and type 2 diabetes is still not confirmed in a Chinese population. The aim of this study was to replicate the associations of GCKR variants with type 2 diabetes and related traits found in Caucasian populations in a Han Chinese population and to identify the potential mechanisms underlying these associations.

## Methods

## Study population

All participants were of Southern Han Chinese ancestry and resided in the Shanghai metropolitan area. We recruited 1118 unrelated type 2 diabetic inpatients from the Endocrinology and Metabolism Department of Zhongshan Hospital, Fudan University, Shanghai, China. All diabetic patients met the 1999 WHO criteria for diabetes [15], had been diagnosed after the age of 29 years, and were treated with oral hypoglycemic agents and/or insulin. The 1161 unrelated non-diabetic control

participants were recruited from people undergoing health examinations in Zhongshan Hospital, were older than 40 years, and had a fasting plasma glucose < 5.6 mmol/l.

Written informed consent was obtained from all participants and the study was approved by the ethnic committee of Zhongshan Hospital, Fudan University, Shanghai, China.

#### Clinical measurements

Both the diabetic patients and the controls were extensively phenotyped for anthropometric and biochemical traits related to glucose metabolism. The phenotypes assessed in our study include height, weight, waist circumference, blood pressure, fasting glucose, total cholesterol, triglyceride, high density lipoprotein cholesterol (HDL-C), and low density lipoprotein cholesterol (LDL-C). BMI was calculated as weight (kg)/height<sup>2</sup> (m<sup>2</sup>). In a subgroup of diabetic patients (n = 664), potential beta cell function was determined using intravenous arginine stimulation tests under fasting conditions. After taking a baseline blood sample, a 10% (wt/vol.) solution of arginine hydrochloride (5 g) was injected intravenously for 30-45 s. The end of the injection period was designated as time zero, after which samples were taken at 2, 4 and 6 min. The acute insulin response (AIR) to arginine was calculated as the mean of the insulin levels in the postinjection samples minus the insulin level in the prestimulus sample. The acute C-peptide response (ACPR) to arginine was calculated in the same way using sampled C-peptide levels.

#### Genotyping

We selected tag single nucleotide polymorphisms (SNPs) across the region of the *GCKR* gene (include 20 kb upstream and 9 kb downstream of the gene) from the HapMap Phase II, using the pairwise tagging model.

The pairwise tagging algorithm was developed by Carlson et al. and has been described previously [16]. In brief, the algorithm is based on the r<sup>2</sup> LD statistic and is comprised of several steps [16]. Starting with all SNPs above the specified minor allele frequency (MAF) threshold in the candidate gene region, the single SNP exceeding the specified r<sup>2</sup> threshold with the maximum number of other SNPs above the MAF threshold is identified [16]. This maximally informative SNP and all associated SNPs are grouped as a bin of associated sites [16]. Any SNP exceeding the threshold r<sup>2</sup> with all other SNPs in the bin is specified as a tag SNP for the bin [16]. Thus, one or more SNPs within a bin are specified as "tag SNPs" and only one tag SNP would need to be genotyped per bin. The binning process is iterated, analyzing all as-yet-unbinned SNPs at each round, until all sites exceeding the MAF

threshold are binned [16]. Thus, the maximally informative set of common SNPs (tag SNPs) is selected and is to be assayed in candidate gene association studies [16]. All polymorphisms above a specified frequency threshold either are directly assayed or exceed a specified threshold level of r<sup>2</sup> with an assayed polymorphism (tag SNP) [16].

The selection criteria used in our study were an r<sup>2</sup> > 0.8 and a minor allele frequency > 0.05. Finally, four tag SNPs (rs8179206, rs2293572, rs3817588 and rs780094) were selected and genotyped. The genotyping was performed by matrix-assisted laser desorption/ionization time-of-flight mass spectroscopy using a MassARRAY platform (MassARRAY Compact Analyzer, Sequenom, San Diego, CA, USA).

#### Statistical analysis

Continuous variables are expressed as the means ± SEM. Comparisons between groups were performed with T testing and  $\chi^2$  testing for normally distributed continuous and categorical variables, respectively. Deviations from the Hardy-Weinberg equilibrium were assessed by means of  $\chi^2$  testing. SNPs that were not in Hardy-Weinberg equilibrium were excluded from further analysis. Pairwise linkage disequilibrium including D' and r<sup>2</sup> was estimated using Haploview. Haplotypes estimating from the population genotype data were performed in Haplo. Stats (R2.8.1). We did allelic analysis for the association of GCKR polymorphisms with type 2 diabetes using logistic regression. We did genotypic analysis for the association of GCKR polymorphisms with quantitative traits using a general linear model, assuming an additive model. We tested the association of haplotypes with type 2 diabetes and quantitative traits by using logistic regression and a general linear model. Non-normally distributed values were logtransformed before analysis. All models were adjusted for year of birth and sex. Additional models were adjusted for BMI. Analysis was performed using SPSS software version 13.0. We used Bonferroni correction for multiple testing.

#### Results

#### Baseline characteristics

The baseline characteristics of participants in this study are presented in Table 1. Of 2279 participants, 1118 were type 2 diabetes patients and 1161 were non-diabetic controls. Diabetic cases were older and had higher BMI, waist circumference, fasting glucose and triglyceride levels, but lower cholesterol concentrations than non-diabetic controls. There was no significant difference in the distribution of sex between diabetic cases and non-diabetic controls (p = 0.34).

## Associations of GCKR polymorphisms with type 2 diabetes

Overall, four SNPs (rs8179206, rs2293572, rs3817588 and rs780094) were selected and genotyped in the present study. The call rates of rs8179206, rs2293572, rs3817588 and rs780094 were 98.5%, 98.0%, 96.6% and 98.5%, respectively. The concordant rates of all SNPs based on 120 duplicates were 100%.

Rs8179206, rs2293572, rs3817588 and rs780094 were in Hardy-Weinberg equilibrium in the total population, diabetic cases and non-diabetic controls (Table 2). The G allele of rs3817588 was significantly associated with an increased risk of type 2 diabetes after adjustment for year of birth and sex (OR = 1.21, 95% CI 1.06-1.39, p = 0.004) (Table 3). In addition, the G allele of rs780094 was significantly associated with an increased risk of type 2 diabetes after adjustment for year of birth and sex (OR = 1.19, 95% CI 1.05-1.34, p = 0.005) (Table 3).

Table 1 Baseline characteristics of all genotyped participants, diabetic cases and non-diabetic controls

Characteristic	All participants (n = 2279)	Diabetic cases $(n = 1118)$	Non-diabetic controls $(n = 1161)$	P value*	
Age (years)	58.3 ± 0.24	60.2 ± 0.37	56.5 ± 0.32	< 0.001	
Men (%)	43.6	44.6	42.7	0.34	
BMI (kg/m²)	$24.0 \pm 0.07$	$24.4 \pm 0.11$	$23.5 \pm 0.09$	< 0.001	
Waist circumference (cm)	85.6 ± 0.45	90.5 ± 0.81	80.5 ± 0.28	< 0.001	
Fasting glucose (mmol/l)	$6.30 \pm 0.01$	$8.17 \pm 0.04$	$4.86 \pm 0.01$	< 0.001	
Total cholesterol (mmol/l)+	$4.76 \pm 0.03$	$4.43 \pm 0.04$	$5.10 \pm 0.03$	< 0.001	
Triglyceride (mmol/l)+	$1.54 \pm 0.02$	$1.64 \pm 0.03$	$1.46 \pm 0.02$	< 0.001	
HDL-C (mmol/l)*	$1.24 \pm 0.01$	$1.16 \pm 0.01$	$1.32 \pm 0.01$	< 0.001	
LDL-C (mmol/l)+	$2.69 \pm 0.02$	$2.40 \pm 0.03$	$3.00 \pm 0.02$	< 0.001	
Systolic BP (mmol/l)	$126.8 \pm 0.38$	$134.2 \pm 0.53$	119.4 ± 0.45	< 0.001	
Diastolic BP (mmol/l)	$79.4 \pm 0.19$	$80.7 \pm 0.28$	$78.0 \pm 0.26$	< 0.001	

Continuous data are expressed as the means  $\pm$  SEM

<sup>&</sup>lt;sup>+</sup>Variables were log-transformed before statistical analysis; numbers in the table were back-transformed

<sup>\*</sup>p value for comparison between cases and controls

Table 2 Characteristics of SNPs genotyped in GCKR

	SNP identification	Chromosome Position	Relation to the gene	Major allele	Minor allele	MAF	HWE P value
1	rs8179206	22573946	Exon	Α	G	0.03	0.76
2	rs2293572	27582281	Intron	G	C	0.14	0.85
3	rs3817588	27584716	Intron	Α	G	0.30	0.10
4	rs780094	27594741	Intron	Α	G	0.45	0.90

The associations remained significant after additional adjustment for BMI (Table 3). The associations of rs8179206 and rs2293572 with type 2 diabetes were not significant (OR = 1.18, 95% CI 0.85-1.65, p = 0.32 and OR = 1.04, 95% CI 0.88-1.23, p = 0.64, respectively) (Table 3).

# Associations of GCKR polymorphisms with quantitative traits in non-diabetic controls

In the non-diabetic controls, the GG and AG carriers of rs3817588 were nominally associated with a lower plasma triglyceride level compared with the AA carriers after adjustment for year of birth, sex and BMI (p = 0.0003 and p = 0.02, respectively), and the trend was in the same direction (p for trend = 0.00004) (Table 4). The association of rs3817588 with plasma triglyceride level was still significant after correction for multiple testing. The GG carriers of rs780094 were nominally associated with a lower plasma triglyceride level compared with the AA carriers, after adjustment for year of birth, sex and BMI (p = 0.01), and the trend was in the same direction (p for trend = 0.03) (Table 4). However, the association of rs780094 with plasma triglyceride level was not significant after correction for multiple testing. The associations of rs8179206 and rs2293572 with plasma triglyceride level were not significant (Table 4). The GG carriers of rs3817588 were nominally associated with a higher waist circumference compared with the AA carriers, after adjustment for year of birth and sex (p = 0.01), and the trend was in the same direction (p for trend = 0.04) (Table 4). The association was not significant after correction for multiple testing. The associations of other polymorphisms with waist circumference were not significant (Table 4). None of the four polymorphisms showed a significant association with BMI, fasting plasma glucose, total cholesterol, HDL-C, LDL-C, systolic blood pressure or diastolic blood pressure (Table 4).

# Associations of *GCKR* polymorphisms with AIR and ACPR in diabetic cases

A subgroup of diabetic cases (n = 664) was classified into 4 groups according to the duration of diabetes. The range of diabetes duration was 1 month to 40 years in this subgroup of diabetic patients. In the third quartile subgroup with a diabetic duration of 8-11 years, the GG carriers of rs780094 were nominally associated with lower levels of AIR and ACPR compared with the AA carriers, after adjustment for year of birth, sex and BMI (p = 0.02 and 0.03, respectively), and the trend was in

Table 3 Allelic distribution of GCKR polymorphisms and association with type 2 diabetes

Alleles	Non-DM (%)	DM (%)	Odds Ratio1 <sup>a</sup>	95% CI	P value	Odds Ratio2 <sup>b</sup>	95% CI	P value
rs8179206								
Α	97.0	96.6	1	(Ref.)		1	(Ref.)	
G	3.0	3.4	1.18	0.85-1.65	0.32	1.29	0.91-1.82	0.15
rs2293572								
G	85.9	85.6	1	(Ref.)		1	(Ref.)	
C	14.1	14.4	1.04	0.88-1.23	0.64	1.04	0.88-1.24	0.64
rs3817588								
Α	72.1	68.0	1	(Ref.)		1	(Ref.)	
G	27.9	32.0	1.21	1.06-1.39	0.004*	1.24	1.08-1.43	0.002*
rs780094								
Α	56.6	52.6	1	(Ref.)		1	(Ref.)	
G	43.4	47.4	1.19	1.05-1.34	0.005*	1.22	1.07-1.38	0.002*

<sup>&</sup>lt;sup>a</sup> Adjusted for year of birth and sex

<sup>&</sup>lt;sup>b</sup> Adjusted for year of birth, sex and BMI

<sup>\*</sup>Significant after Bonferroni correction (p < 0.0125 (0.05/4) was the corrected statistically significant level in association analysis between the individual SNP and type 2 diabetes)

The major allele was used as a reference when comparing the risk of diabetes between alleles.

Table 4 Quantitative traits stratified according to GCKR genotypes in non-diabetic controls

	n	BMI (kg/m²)	Waist circumference (cm)	Fasting glucose (mmol/l)	Total cholesterol* (mmol/l)	Triglyceride* (mmol/l)	HDL-C* (mmol/l)	LDL-C* (mmol/l)	Systolic BP (mmHg)	Diastolic BP (mmHg)
rs8179206										
AA	1076	23.50 ± 0.09	80.50 ± 0.29	$4.86 \pm 0.01$	$5.11 \pm 0.03$	$1.45 \pm 0.02$	$1.32 \pm 0.01$	$3.01 \pm 0.03$	119.14 ± 0.46	77.95 ± 0.27
AG	68	23.33 ± 0.39	81.48 ± 1.23	$4.86 \pm 0.05$	$5.04 \pm 0.11$	$1.52 \pm 0.09$	$1.30 \pm 0.03$	$2.95 \pm 0.10$	122.28 ± 1.99	79.55 ± 1.11
GG <sup>\$</sup>	2	19.47 ± /	68.00 ± /	4.10 ± /	$3.80 \pm /$	0.90 ± /	1.16 ± /	2.23 ± /	110.00 ± /	70.00 ± /
P value <sup>a#</sup>		0.36	0.33	0.19	0.27	0.51	0.61	0.53	0.45	0.76
P value <sup>b#</sup>				0.15	0.31	0.49	0.24	0.64	0.35	0.72
rs2293572										
GG	842	23.53 ± 0.10	$80.60 \pm 0.32$	$4.85 \pm 0.01$	$5.09 \pm 0.03$	$1.44 \pm 0.02$	$1.32 \pm 0.01$	$3.00 \pm 0.03$	119.32 ± 0.53	$78.13 \pm 0.31$
GC	276	23.37 ± 0.17	80.22 ± 0.58	$4.87 \pm 0.02$	$5.16 \pm 0.06$	$1.51 \pm 0.04$	$1.31 \pm 0.02$	$3.03 \pm 0.05$	118.96 ± 0.90	$77.54 \pm 0.50$
CC	23	$24.03 \pm 0.53$	$80.00 \pm 1.85$	$4.98 \pm 0.06$	$5.39 \pm 0.19$	$1.53 \pm 0.15$	$1.39 \pm 0.06$	$3.12 \pm 0.17$	121.24 ± 1.95	78.96 ± 1.29
P value <sup>a#</sup>		0.48	0.63	0.18	0.17	0.28	0.54	0.74	0.14	0.24
P value <sup>b#</sup>				0.14	0.17	0.15	0.37	0.73	0.19	0.44
rs3817588										
AA	598	$23.35 \pm 0.12$	$79.88 \pm 0.40$	$4.85 \pm 0.02$	$5.13 \pm 0.04$	$1.50 \pm 0.03$	$1.32 \pm 0.01$	$3.02 \pm 0.04$	$119.27 \pm 0.66$	$77.79 \pm 0.38$
AG	423	23.52 ± 0.15	$81.14 \pm 0.47$	$4.87 \pm 0.02$	$5.08 \pm 0.05$	$1.41 \pm 0.03$	$1.33 \pm 0.02$	$3.00 \pm 0.04$	$118.93 \pm 0.76$	78.12 ± 0.45
GG	103	$23.97 \pm 0.30$	82.13 ± 1.01	$4.84 \pm 0.04$	$4.99 \pm 0.10$	$1.26 \pm 0.05$	$1.32 \pm 0.03$	$2.95 \pm 0.08$	119.58 ± 1.68	$77.77 \pm 0.94$
P value <sup>a#</sup>		0.15	0.04	0.84	0.35	0.001	0.78	0.82	0.59	0.76
P value <sup>b#</sup>				0.81	0.29	0.00004	0.70	0.84	0.39	0.63
rs780094										
AA	368	23.22 ± 0.16	$79.66 \pm 0.50$	$4.84 \pm 0.02$	$5.12 \pm 0.05$	$1.49 \pm 0.04$	$1.32 \pm 0.02$	$3.02 \pm 0.05$	119.13 ± 0.87	$77.83 \pm 0.50$
AG	558	23.59 ± 0.13	$80.80 \pm 0.42$	$4.86 \pm 0.02$	$5.13 \pm 0.04$	$1.44 \pm 0.03$	$1.32 \pm 0.01$	$3.02 \pm 0.04$	$119.73 \pm 0.70$	$78.40 \pm 0.39$
GG	218	23.67 ± 0.20	$81.34 \pm 0.70$	$4.85 \pm 0.03$	$5.00 \pm 0.07$	$1.35 \pm 0.04$	$1.32 \pm 0.02$	$2.92 \pm 0.06$	119.15 ± 1.04	$77.33 \pm 0.62$
P value <sup>a#</sup>		0.15	0.16	0.74	0.32	0.06	0.79	0.40	0.96	0.24
P value <sup>b#</sup>				0.63	0.31	0.03	0.30	0.43	0.99	0.18

Data are expressed as the means  $\pm$  SEM

<sup>&</sup>lt;sup>a</sup> Adjusted for year of birth and sex

<sup>&</sup>lt;sup>b</sup> Adjusted for year of birth, sex and BMI

<sup>\*</sup> Variables were log-transformed before statistical analysis; numbers in the table were back-transformed

<sup>&</sup>lt;sup>#</sup> p for trend

<sup>\$</sup> This group had too few subjects and no SEM was calculated

<sup>&</sup>lt;sup>&</sup>Significant after Bonferroni correction (p < 0.0014 (0.05/36) was the corrected statistically significant level in association analysis between the individual SNP and quantitative traits in the control group)

the same direction (p for trend = 0.03 and 0.09, respectively) (Figure 1). In the fourth quartile subgroup with a diabetic duration over 11 years, the GG carriers of rs3817588 were nominally associated with lower levels of AIR and ACPR compared with the AA carriers, after adjustment for year of birth, sex and BMI (p = 0.008 and 0.01, respectively), and the trend was in the same direction (p for trend = 0.03 and 0.03, respectively) (Figure 1). However, these associations were no longer significant after correction for multiple testing.

#### Associations of GCKR haplotypes with type 2 diabetes

The haplotype block was constructed for the four SNPs (rs8179206, rs2293572, rs3817588 and rs780094) in *GCKR*. All four SNPs fell into one block (Additional file 1, Figure S1). The block was associated with type 2 diabetes after adjustment for year of birth and sex (the global p value =  $2.7 \times 10^{-5}$ ) (Additional file 1, Table S1). The haplotypes AGGG and GGGG were associated with an increased risk of type 2 diabetes (OR = 1.18, 95% CI 1.02-1.37, p = 0.02 and OR = 2.08, 95% CI 1.60-2.71, p =  $5.9 \times 10^{-8}$ , respectively) (Additional file 1, Table S1). The associations remained similar after additional adjustment for BMI (Additional file 1, Table S1).

# Associations of *GCKR* haplotypes with quantitative traits in non-diabetic controls

In the non-diabetic controls, the haplotype block was nominally associated with plasma triglyceride level adjusted for year of birth, sex and BMI (the global p value = 0.0001) (Additional file 1, Table S2). The haplotype AGGG was associated with a lower plasma triglyceride level adjusted for year of birth, sex and BMI (p =  $2.2 \times 10^{-5}$ ). The association remained significant after correction for multiple testing. BMI, waist circumference, fasting plasma glucose, total cholesterol, HDL-C, LDL-C, systolic blood pressure and diastolic blood pressure were not significantly different between haplotypes (Additional file 1, Table S2).

#### Discussion

In line with previous studies, our study confirmed the opposite effects of *GCKR* variants on glucose and triglyceride concentrations. Our data showed that the *GCKR* rs780094 G allele and rs3817588 G allele were associated with an increased risk of type 2 diabetes in Han Chinese individuals. The G alleles were also nominally associated with a lower fasting triglyceride level. Moreover, the association of rs3817588 with fasting triglyceride level was still significant after correction for multiple testing. The associations of *GCKR* rs780094 with type 2 diabetes and triglyceride level have been replicated in many studies of different ethnic populations since the Diabetes Genetics Initiative genome-wide association

study [7-11]. Our study confirmed this association again in a Han Chinese population. Our study was the first study which adopted a tagging strategy and selected tag SNPs of *GCKR* including 20 kb upstream and 9 kb downstream of the gene for studying. We demonstrated that another polymorphism rs3817588 in *GCKR* affected glucose and lipid metabolism in a similar way as rs780094, which was not reported in the previous studies.

In the present study, the G allele of GCKR rs780094 was associated with higher odds of type 2 diabetes (OR = 1.19). The effect size was similar to that observed in another study of a Han Chinese population [12]. In the studies of populations of European descent, the G allele of rs780094 was also associated with a higher odds of diabetes, but the effect size was much smaller (OR = 1.06-1.08) [8,10,11]. The frequency of the rs780094 G allele is substantially lower in Han Chinese (43%) than in White Europeans (65%). The difference in genetic background between different ethnic groups may explain the discrepancy between effect sizes in Han Chinese and European populations. The G allele of GCKR rs3817588 was associated with higher odds of type 2 diabetes (OR = 1.21). For both polymorphisms, the G risk alleles for diabetes were nominally associated with a lower triglyceride level. The mechanism by which the GCKR variants lead to type 2 diabetes and protect against dyslipidemia remains to be determined. A potential explanation is that GCK regulation by GCKR is altered in the liver, which leads indirectly to decreased GCK activity [17]. Decreased GCK activity was associated with decreased glucose utilization in the liver [17]. With decreased glucose utilization and glycolytic flux, GCK, phosphofructokinase, and fatty acid synthase are downregulated, whereas phosphoenolpyruvate carboxykinase and glucose-6-phosphatase are upregulated [17]. These changes increase hepatic glucose output, lower malonyl-CoA concentration and inhibit de novo lipogenesis and VLDL triglyceride production [17].

We next investigated whether the *GCKR* variants were associated with beta cell function as determined by an arginine stimulation test in diabetic patients. We found that in groups with a relatively long diabetic duration, *GCKR* variants were associated with AIR and ACPR. The carriers of the G alleles of rs780094 and rs3817588 had lower values for AIR and ACPR than the AA homozygotes. These findings suggested that the *GCKR* variants probably contribute to diabetes susceptibility by impairing beta cell function, although the associations of the *GCKR* variants with AIR and ACPR became nonsignificant after correction for multiple testing. A study in a Han Chinese population found that rs780094 was associated with beta cell function as estimated by HOMA-B, which was consistent with our findings [12].

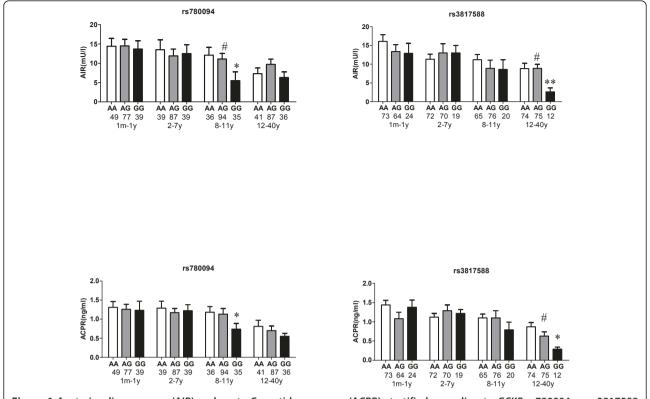


Figure 1 Acute insulin response (AIR) and acute C-peptide response (ACPR) stratified according to GCKR rs780094 or rs3817588 genotypes by quartile of diabetic duration. \* p < 0.05 compared with homozygote of major allele (AA) \*\* p < 0.01 compared with homozygote of major allele (AA) # p for trend < 0.05 All p values were not significant after Bonferroni correction (p < 0.0016 (0.05/32) was used as Bonferroni corrected statistically significant level in association analysis between the individual SNP and AIR or ACPR) The figures below the genotypes indicate the number of diabetic cases for each genotype group.

We also did haplotype analysis and found that all four SNPs exhibited moderate to strong LD in terms of D' and fell into one block. The block was associated with type 2 diabetes after adjustment for year of birth, sex and BMI. The AGGG haplotype and the GGGG haplotype were associated with an increased risk of type 2 diabetes. Both haplotypes carried the G risk alleles of rs3817588 and rs780094. However, the ACAG haplotype which carried the G risk allele of rs780094 did not show any association with type 2 diabetes. This would suggest that the effect of the rs780094 G allele on the risk of type 2 diabetes was due to its LD with rs3817588. The difference of the AGGG haplotype and the GGGG haplotype was at the locus of rs8179206. However, the GGGG haplotype was associated with a 2.08 times risk of diabetes which was much higher than that of the AGGG haplotype (OR = 1.18), suggesting that the G alleles of rs8179206 and rs3817588 had synergistic effect on the development of type 2 diabetes. Rs8179206 G allele also contributed to the risk of type 2 diabetes although it was not associated with diabetes in single locus analysis.

Rs3817588 and rs780094 are located in introns of GCKR. Based on the current data, we cannot confirm whether they are in the splicing site or the transcription factor binding site of the gene. We therefore assume them to be linked with one or more functional variants within the GCKR gene or its regulatory regions. Rs780094 is tightly linked with rs1260326 (HapMap CEU  $r^2 = 0.93$ , CHB  $r^2 = 0.82$ ), a non-synonymous variant in GCKR associated with type 2 diabetes and triglyceride level [9,10,12]. A functional study showed that GCKR rs1260326 was associated with fasting plasma glucose and triglyceride levels, and this effect was mediated through regulating the activity of GCK in liver [17]. We did not genotype rs1260326 in the current study because of the fact that it is in strong LD with rs780094 and represents the same information as rs780094, which was demonstrated by a previous study in a Han Chinese population [12]. Although there is no evidence that rs3817588 is linked with any functional variant now, future fine mapping and resequencing of the GCKR gene may detect such functional variants.

Our study had some limitations. Firstly, we did not investigate gene-environment interactions. Because both genetic variants and environmental factors contribute to type 2 diabetes, and adverse environmental factors (high-caloric diets, physical inactivity, etc.) have important influence on the development of diabetes, the elucidation of gene-environment interactions should not be overlooked in future studies. Secondly, we did not do a functional study of rs3817588. Further functional study is needed to determine whether rs3817588 is associated with selective splicing of mRNA or the binding of transcription factors and affects expression level of protein ultimately.

### **Conclusions**

We demonstrated that the rs3817588 A/G polymorphism of the *GCKR* gene was associated with type 2 diabetes and plasma triglyceride level in the Han Chinese population.

#### **Additional material**

Additional file 1: Table S1 Association of *GCKR* haplotypes with type 2 diabetes. Table S2 Quantitative traits stratified according to *GCKR* haplotypes in non-diabetic controls Figure S1 Haploview-generated linkage disequilibrium (LD) map and blocks of the 4 SNPs at the *GCKR* locus

#### Abbreviations

GCK: glucokinase; GCKR: glucokinase regulatory protein; HOMA-IR: homeostasis model assessment of insulin resistance; HOMA-B: homeostasis model assessment of beta cell function; IFG: impaired fasting glucose; AIR: acute insulin response; ACPR: acute C-peptide response; SNP: single nucleotide polymorphism; MAF: minor allele frequency; LD: linkage disequilibrium; SEM: standard error of mean; BMI: body mass index; CI: confidence interval; OR: odds ratio; HDL-C: high density lipoprotein cholesterol; LDL-C: low density lipoprotein cholesterol; BP: blood pressure; DM: diabetes; HWE: Hardy-Weinberg equilibrium.

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## Authors' contributions

YL participated in the design of the study, carried out the SNP genotyping and the statistical analysis of the genotype data, and drafted the manuscript. XG contributed to the design and coordination of the study, to the statistical analysis, interpreted the findings and drafted the manuscript. XL, QG, HC and DL participated in the design of the study and the SNP genotyping. All authors read and approved the final manuscript.

#### Competing interests

The authors declare that they have no competing interests.

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#### References

- Matschinsky FM: Banting lecture 1995. A lesson in metabolic regulation inspired by the glucokinase glucose sensor paradigm. *Diabetes* 1996, 45(2):223-241.
- Van Schaftingen E: A protein from rat liver confers to glucokinase the property of being antagonistically regulated by fructose 6-phosphate and fructose 1-phosphate. Eur J Biochem 1989, 179(1):179-184.
- Chu CA, Fujimoto Y, İgawa K, Grimsby J, Grippo JF, Magnuson MA, Cherrington AD, Shiota M: Rapid translocation of hepatic glucokinase in response to intraduodenal glucose infusion and changes in plasma glucose and insulin in conscious rats. Am J Physiol Gastrointest Liver Physiol 2004, 286(4):G627-G634.
- Farrelly D, Brown KS, Tieman A, Ren J, Lira SA, Hagan D, Gregg R, Mookhtiar KA, Hariharan N: Mice mutant for glucokinase regulatory protein exhibit decreased liver glucokinase: a sequestration mechanism in metabolic regulation. Proc Natl Acad Sci USA 1999, 96(25):14511-14516.
- Grimsby J, Coffey JW, Dvorozniak MT, Magram J, Li G, Matschinsky FM, Shiota C, Kaur S, Magnuson MA, Grippo JF: Characterization of glucokinase regulatory protein-deficient mice. J Biol Chem 2000, 275(11):7826-7831.
- Slosberg ED, Desai UJ, Fanelli B, St Denny I, Connelly S, Kaleko M, Boettcher BR, Caplan SL: Treatment of type 2 diabetes by adenoviralmediated overexpression of the glucokinase regulatory protein. *Diabetes* 2001, 50(8):1813-1820.
- 7. Diabetes Genetics Initiative of Broad Institute of Harvard and MIT, Lund University, and Novartis Institutes of BioMedical Research, Saxena R, Voight BF, Lyssenko V, Burtt NP, de Bakker PI, Chen H, Roix JJ, Kathiresan S, Hirschhorn JN, Daly MJ, Hughes TE, Groop L, Altshuler D, Almgren P, Florez JC, Meyer J, Ardlie K, Bengtsson Boström K, Isomaa B, Lettre G, Lindblad U, Lyon HN, Melander O, Newton-Cheh C, Nilsson P, Orho-Melander M, Råstam L, Speliotes EK, Taskinen MR, Tuomi T, Guiducci C, Berglund A, Carlson J, Gianniny L, Hackett R, Hall L, Holmkvist J, Laurila E, Sjögren M, Sterner M, Surti A, Svensson M, Svensson M, Tewhey R, Blumenstiel B, Parkin M, Defelice M, Barry R, Brodeur W, Camarata J, Chia N, Fava M, Gibbons J, Handsaker B, Healy C, Nguyen K, Gates C, Sougnez C, Gage D, Nizzari M, Gabriel SB, Chirn GW, Ma Q, Parikh H, Richardson D, Ricke D, Purcell S: Genome-wide association analysis identifies loci for type 2 diabetes and triglyceride levels. Science 2007, 316(5829):1331-1336.
- Sparsø T, Andersen G, Nielsen T, Burgdorf KS, Gjesing AP, Nielsen AL, Albrechtsen A, Rasmussen SS, Jørgensen T, Borch-Johnsen K, Sandbaek A, Lauritzen T, Madsbad S, Hansen T, Pedersen O: The GCKR rs780094 polymorphism is associated with elevated fasting serum triacylglycerol, reduced fasting and OGTT-related insulinaemia, and reduced risk of type 2 diabetes. *Diabetologia* 2008, 51(1):70-75.
- Vaxillaire M, Cavalcanti-Proença C, Dechaume A, Tichet J, Marre M, Balkau B, Froguel P: DESIR Study Group: The common P446L polymorphism in GCKR inversely modulates fasting glucose and triglyceride levels and reduces type 2 diabetes risk in the DESIR prospective general French population. *Diabetes* 2008, 57(8):2253-2257.
- 10. Orho-Melander M, Melander O, Guiducci C, Perez-Martinez P, Corella D, Roos C, Tewhey R, Rieder MJ, Hall J, Abecasis G, Tai ES, Welch C, Arnett DK, Lyssenko V, Lindholm E, Saxena R, de Bakker Pl, Burtt N, Voight BF, Hirschhorn JN, Tucker KL, Hedner T, Tuomi T, Isomaa B, Eriksson KF, Taskinen MR, Wahlstrand B, Hughes TE, Parnell LD, Lai CQ, Berglund G, Peltonen L, Vartiainen E, Jousilahti P, Havulinna AS, Salomaa V, Nilsson P, Groop L, Altshuler D, Ordovas JM, Kathiresan S: Common missense variant in the glucokinase regulatory protein gene is associated with increased plasma triglyceride and C-reactive protein but lower fasting glucose concentrations. Diabetes 2008, 57:3112-3121.
- Dupuis J, Langenberg C, Prokopenko I, Saxena R, Soranzo N, Jackson AU, Wheeler E, Glazer NL, Bouatia-Naji N, Gloyn AL, Lindgren CM, Mägi R, Morris AP, Randall J, Johnson T, Elliott P, Rybin D, Thorleifsson G, Steinthorsdottir V, Henneman P, Grallert H, Dehghan A, Hottenga JJ, Franklin CS, Navarro P, Song K, Goel A, Perry JR, Egan JM, Lajunen T,

Grarup N, Sparsø T, Doney A, Voight BF, Stringham HM, Li M, Kanoni S, Shrader P, Cavalcanti-Proenca C, Kumari M, Qi L, Timpson NJ, Gieger C, Zabena C, Rocheleau G, Ingelsson E, An P, O'Connell J, Luan J, Elliott A, McCarroll SA, Payne F, Roccasecca RM, Pattou F, Sethupathy P, Ardlie K, Ariyurek Y, Balkau B, Barter P, Beilby JP, Ben-Shlomo Y, Benediktsson R, Bennett AJ, Bergmann S, Bochud M, Boerwinkle E, Bonnefond A, Bonnycastle LL, Borch-Johnsen K, Böttcher Y, Brunner E, Bumpstead SJ, Charpentier G, Chen YD, Chines P, Clarke R, Coin LJ, Cooper MN, Cornelis M, Crawford G, Crisponi L, Day IN, de Geus EJ, Delplanque J, Dina C, Erdos MR, Fedson AC, Fischer-Rosinsky A, Forouhi NG, Fox CS, Frants R, Franzosi MG, Galan P, Goodarzi MO, Graessler J, Groves CJ, Grundy S, Gwilliam R, Gyllensten U, Hadjadj S, Hallmans G, Hammond N, Han X, Hartikainen AL, Hassanali N, Hayward C, Heath SC, Hercberg S, Herder C, Hicks AA, Hillman DR, Hingorani AD, Hofman A, Hui J, Hung J, Isomaa B, Johnson PR, Jørgensen T, Jula A, Kaakinen M, Kaprio J, Kesaniemi YA, Kivimaki M, Knight B, Koskinen S, Kovacs P, Kyvik KO, Lathrop GM, Lawlor DA, Le Bacquer O, Lecoeur C, Li Y, Lyssenko V, Mahley R, Mangino M, Manning AK, Martínez-Larrad MT, McAteer JB, McCulloch LJ, McPherson R, Meisinger C, Melzer D, Meyre D, Mitchell BD, Morken MA Mukheriee S Naitza S Narisu N Neville M L Oostra BA Orrù M Pakyz R, Palmer CN, Paolisso G, Pattaro C, Pearson D, Peden JF, Pedersen NL, Perola M, Pfeiffer AF, Pichler I, Polasek O, Posthuma D, Potter SC, Pouta A, Province MA, Psaty BM, Rathmann W, Rayner NW, Rice K, Ripatti S, Rivadeneira F, Roden M, Rolandsson O, Sandbaek A, Sandhu M. Sanna S. Saver AA, Scheet P. Scott LJ, Seedorf U. Sharp SJ. Shields B, Sigurethsson G, Sijbrands EJ, Silveira A, Simpson L, Singleton A, Smith NL, Sovio U, Swift A, Syddall H, Syvänen AC, Tanaka T, Thorand B, Tichet J, Tönjes A, Tuomi T, Uitterlinden AG, van Dijk KW, van Hoek M, Varma D, Visvikis-Siest S, Vitart V, Vogelzangs N, Waeber G, Wagner PJ, Walley A, Walters GB, Ward KL, Watkins H, Weedon MN, Wild SH, Willemsen G, Witteman JC, Yarnell JW, Zeggini E, Zelenika D, Zethelius B, Zhai G, Zhao JH, Zillikens MC, DIAGRAM Consortium, GIANT Consortium, Global BPgen Consortium, Borecki IB, Loos RJ, Meneton P, Magnusson PK, Nathan DM, Williams GH, Hattersley AT, Silander K, Salomaa V, Smith GD, Bornstein SR, Schwarz P, Spranger J, Karpe F, Shuldiner AR, Cooper C, Dedoussis GV, Serrano-Ríos M, Morris AD, Lind L, Palmer LJ, Hu FB, Franks PW, Ebrahim S, Marmot M, Kao WH, Pankow JS, Sampson MJ, Kuusisto J, Laakso M, Hansen T, Pedersen O, Pramstaller PP, Wichmann HE, Illig T, Rudan I, Wright AF, Stumvoll M, Campbell H, Wilson JF, Anders Hamsten on behalf of Procardis Consortium, MAGIC investigators, Bergman RN, Buchanan TA, Collins FS, Mohlke KL, Tuomilehto J, Valle TT, Altshuler D, Rotter JI, Siscovick DS, Penninx BW, Boomsma DI, Deloukas P, Spector TD, Frayling TM, Ferrucci L, Kong A, Thorsteinsdottir U, Stefansson K, van Duijn CM, Aulchenko YS, Cao A, Scuteri A, Schlessinger D, Uda M, Ruokonen A, Jarvelin MR, Waterworth DM, Vollenweider P, Peltonen L, Mooser V, Abecasis GR, Wareham NJ, Sladek R, Froguel P, Watanabe RM, Meigs JB, Groop L, Boehnke M, McCarthy MI, Florez JC, Barroso I: New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. Nature Genetics 2010, 42(2):105-116.

- Qi Q, Wu Y, Li H, Loos RJ, Hu FB, Sun L, Lu L, Pan A, Liu C, Wu H, Chen L, Yu Z, Lin X: Association of GCKR rs780094, alone or in combination with GCK rs1799884, with type 2 diabetes and related traits in a Han Chinese population. *Diabetologia* 2009, 52(5):834-843.
- Tam CH, Ma RC, So WY, Wang Y, Lam VK, Germer S, Martin M, Chan JC, Ng MC: Interaction effect of genetic polymorphisms in glucokinase (GCK) and glucokinase regulatory protein (GCKR) on metabolic traits in healthy Chinese adults and adolescents. *Diabetes* 2009, 58(3):765-769.
- Wen J, Rönn T, Olsson A, Yang Z, Lu B, Du Y, Groop L, Ling C, Hu R: Investigation of type 2 diabetes risk alleles support CDKN2A/B, CDKAL1, and TCF7L2 as susceptibility genes in a Han Chinese cohort. PLoS One 2010, 5(2):e9153.
- World Health Organization: Definition, diagnosis and classification of diabetes mellitus and its complication. WHO/NCD/NCS 1999, 31-32.
- Carlson CS, Eberle MA, Rieder MJ, Yi Q, Kruglyak L, Nickerson DA: Selecting a maximally informative set of single-nucleotide polymorphisms for association analyses using linkage disequilibrium. Am J Hum Genet 2004, 74(1):106-120.
- 17. Beer NL, Tribble ND, McCulloch LJ, Roos C, Johnson PR, Orho-Melander M, Gloyn AL: The P446L variant in GCKR associated with fasting plasma

glucose and triglyceride levels exerts its effect through increased glucokinase activity in liver. *Hum Mol Genet* 2009, **18(21)**:4081-4088.

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